

# **Mapping and Modelling Landscape-based Soil Fertility Change in Relation to Human Induction**

**Case study: GISHWATI Watershed of the Rwandan highlands**

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# Mapping and Modelling Landscape-based Soil Fertility Change in relation to Human Induction

by

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*Dedicated to my husband Fredrick  
And  
My son Jackson and my daughter Lillian*

# Abstract

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Changes in land use and land cover are central to the study of global environmental change. Among these changes is soil fertility degradation, which has become a major problem for agricultural management in Rwanda. Man as a soil-forming factor has been a difficult issue for pedology in general; explaining changes in soil fertility is but one example.

The main agent causing change in processes controlling soil fertility is generally considered to be human activity. However, the nature and causes of soil fertility change in the complex lithology of Rwandan highlands is currently poorly known. In order to design and implement the national policy 2020 for conservation and restoration of soil fertility, policy makers need a clear and quantitative view of the spatio-temporal pattern of nutrient removal and redistribution, as well as the causes of decline. Existing detailed soil maps and lab analyses are more than 20 years old and poorly reflect the situation since the civil conflict of the 1990's, which resulted in major land use changes.

This study attempts a quantitative assessment of human-induced soil fertility change in the Gishwati watershed. To supplement the 1980's soil map, a supplementary soil sampling, followed by lab analysis was carried out. Factor analysis was used to select most-significant fertility indicators (MSFI). These were used to develop a soil fertility index (SFI) and deterioration index (DI) which was interpolated over the study area for both dates. The soil fertility model was developed and validated using independent validation set. Kriging prediction was then applied to the data using auxiliary variables such soil mapping units and soil predictive components (SPC) generated from improved DEM and derived hydrological indices.

The result of PCA showed that pH, OC, Al, P, K, Ca, and Mg are the most dynamic chemical properties in Rwanda (refer as MSFI), and those can be used to monitor the change in soil fertility over time. One-way ANOVA revealed that all MSFI are highly affected by LUCC with regards to soil type and topography.

By computing the deviation of SFI from natural forest to other land uses, SFI captured the individual MSFI information, and revealed the change of soil fertility over 25 years. The soil fertility deterioration index (DI), which reflects the changes in soil fertility from natural forest conversion to different land uses with respect to soil type, revealed that soil quality in the study area has been significantly reduced in agricultural lands (-31%), pine plantations (-24%) and pasture land (-16%). However, DI showed that volcanic soils are slowly degraded compared acidic soils. With regular cultivation, Eutrandepts and Andaquepts lost only 7% over more than 11 years.

Of the geostatistical analysis including SMUs and SPCs predictors, SFI was the best choice for representing spatial structure of soil fertility change in relation to LUCC. Prediction accuracy ranged from 91% to 93% in the entire Gishwati watershed. Therefore SFI maps differences enabled us to detect early degradation caused by different change in land uses done in different time which could not be easily seen using individual MSFI.

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## List of abbreviations

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ANOVA	Analysis of variance
ASTER	Advanced Space-borne Thermal Emission and Reflection Radiometer
BADC	Belgium Administration for Development Cooperation
CPR	Carte Pedologique du Rwanda
DBF	Database file
DI	Deterioration Index
DEM	Digital elevation map
DN	Digital Number
DTM	Digital Terrain Model
FA	Factor Analysis
ILWIS	Integrated Land and Water Information System software
GLASOD	Global Assessment Degradation
GLM	Generalised Linear Model
KED	Kriging with External Drift
LP	Landscape position
LUCC	Land Use and Land Cover Change
MDS	Minimum Data Set
MPE	Mean Prediction Error
MSFI	Minimum Soil Fertility Indicators
MLC	Maximum Likelihood Classifier
MINITERRE	Ministere of Land, Environment, Forestry, Water and Mines
NUR	National University of Rwanda
NDVI	Normalized Difference Vegetation Index
NMSE	Normalized Mean Square Error
OK	Ordinary Kriging
PCA	Principal Component Analysis
PCFN	Nyungwe Forest Conservation Project
RK	Regression Kriging
RMSPE	Root Mean Square Prediction Error
SCM	Spearman Correlation Matrix
SFI	Soil Fertility Index
SFID	Soil Fertility Index Development
SHP	Shape file
SMU	Soil Mapping Unit
SPC	Soil Predictive Component
SPI	Stream Power Index
STI	Sediment Transport Index
TM	Thematic Mapper
TWI	Topographic Wetness Index
WCS	Wildlife Conservation Society
WLS	Weighted Least Square

# 1. Introduction

## 1.1. Background

Changes in land use and land cover are central to the study of global environmental change including soil fertility degradation, and reflect the rapid population growth in tropics. As a result of increasing demand for firewood, timber, pasture, shelter and food crops, natural land covers, particularly tropical forests, are being degraded or converted to cropland at an alarming rate (Islam and Weil, 2000). Man as soil-forming factor has been a difficult issue in pedology (Hartemink, 2003), whereas many soils in the world have been drastically altered or degraded as a result of human interference (Hartemink, 2003; Jaiyeoba, 2003; Wu and Tiessen, 2002).

Soil fertility degradation by nutrient depletion, mostly caused by erosion but also by removal of nutrients in crops is one of the threats that agricultural systems in Rwanda are facing. Soil erosion is obviously the most visible and sometimes most destructive form, and it has received considerable attention in Rwanda's national policy (MINITERRE/Rwanda, 2003). This threat, in addition to causing on-site loss of topsoil and reducing the productivity of the land, brings about major off-site environmental effects such as flooding and infrastructure damage. Rwanda's relief consists of high mountains, steep-sloped hills and depressions. Highlands are most wet area and water runoff on steep slopes, coupled with the natural fragility of the soil, carries along soils towards valleys and depressions. A big amount is swept along outside Rwanda. On its way, the Akagera River carries along about 30 kg of soil per second. Maximum land loss is estimated at 557 t ha<sup>-1</sup> yr<sup>-1</sup> (MINITERRE/Rwanda, 2003). This affects a big part of territory, particularly fragile ecosystems of mountain regions in the North and in the West; and ends up by causing a reduction of soil fertility and consequently, the loss of land productivity.

To improve degraded soils and restore their productivity, it is necessary to determine the current status, and see whether this degradation can be explained by use of land in local conditions. The purpose of this study is to assess the effect of LUCC on soil fertility at landscape scale. The fundamental question we attempt to answer in this study is: to what extent the change in land use degrades the soil fertility. A related concern is the degree to which land use coupled with complex lithology on hilly landscape can explain the change in soil fertility over 25 years, and how much can be explained by a model of spatial dependence.

The conceptual diagram of changes in soil fertility at watershed level with respect to the major land uses and related management in Rwandan highlands is shown in figure 1-1.

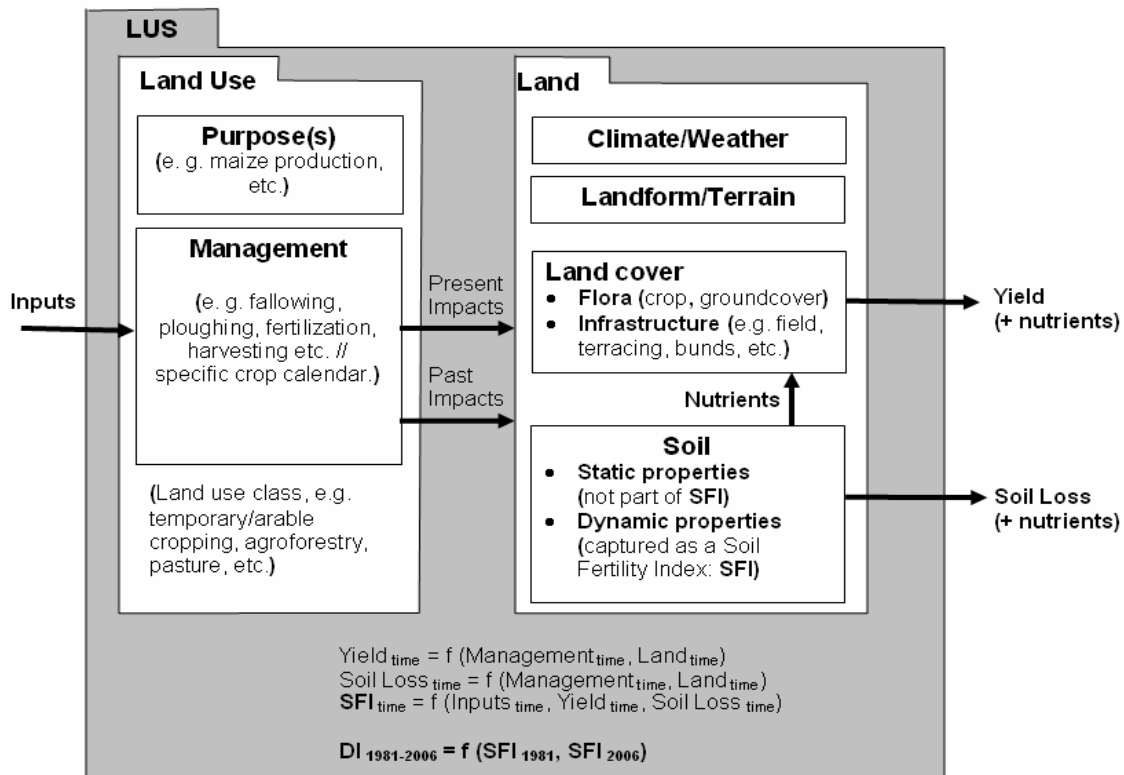


Figure 1-1: Conceptual diagram of soil fertility degradation (DI) in relation to land use system

## 1.2. Research problem

Soil fertility degradation has become a major problem for agricultural management in Rwanda. The main agent causing change in controlling processes is human activity, and a complete explanation of the fertility components cannot be achieved without an understanding of human-induced soil change (Pennock and Veldkamp, 2006) at landscape level.

Land use changes, especially cultivation of deforested land may rapidly diminish soil quality. However, the exact removal of soil fertility in complex lithology of Rwandan highlands is currently poorly known. In order to design and implement the national policy 2020 in conservation and restoration of soil fertility (MINITERRE/Rwanda, 2005), the policy makers need a clear view of the nutrients removal, where this deficiency is located and how much need to be restored. As with accurate information on the distribution of soil type and the soil chemical properties in soil map of Rwanda at scale of 1:50,000 which are more than 20 years, soil change information is needed by today's decision makers for a variety of management goals, including short and long-term productivity, economics, sustainability and environmental quality.

The GISHWATI study area provides an ideal 'laboratory' for assessing soil fertility change, since (1) it was forested until 1994, (2) there was a baseline soil survey done prior to deforestation in 1981, (3)

the area has been deforested since 1995, and due to Agricultural and Settlement activities, it has faced dramatic erosion and changes in soil management, in particular intensive cropping.

### **1.3. Research objectives**

The present study aimed to investigate and map the evolution of soil fertility in Rwandan highland as a result of land use change and related management. The main objective of this research was to quantify the response of soil fertility to human activity in Rwandan highlands at watershed level. The related concerns were to map the spatial distribution of soil fertility using auxiliary variables that were available and to model the temporal evolution of soil fertility within a watershed. To accomplish this, a minimum data set (MDS) for soil fertility changes were developed; and used to determine soil fertility index (SFI) that were used to predict the spatial distribution of changes across different soil types and present land uses of entire Gishwati catchment area.

### **1.4. Research questions**

- 1) What is the Minimum Data Set (MDS) of soil chemical attributes that can be used to assess the landscape-based soil fertility change?
- 2) Is there a significant change in soil fertility over the last 25 years? If so, what is it, and where are the changes most pronounced?
- 3) How can the individual indicators of soil fertility be modeled into an integrative measure of soil fertility and fertility degradation?
- 4) To what extent land use change contributes to soil fertility change at watershed level?
- 5) How successfully can the spatial pattern of soil fertility be predicted in complex lithology of Rwandan highlands?

### **1.5. Hypotheses**

- 1) Soil Nutrients are the most dynamic soil fertility indicators. Those can be used for soil fertility change mapping and modelling.
- 2) The soil fertility indicators have changed significantly over the last 25 years, especially in areas invaded since 1980.
- 3) Soil fertility indicators are modeled into Soil Fertility Index (SFI) and Fertility Deterioration Index (DI) using threshold values of soil properties classes of Rwanda.
- 4) There is significant difference in soil fertility within the same soil forming environment. These differences are highly explained by the type of change in land use.
- 5) The spatial change in soil fertility is fairly mapped using Kriging methods that include soil forming factors in the model of spatial dependence than using only the target variable.

## 1.6. Research approach

Study on how LUCC affects the Soil fertility must involve the response of the soil fertility indicators to LUCC. Numerous models have been adopted to estimate the impact of LUCC on natural resources.

In fact, all the soil properties are not equally affected by the LUCC in space and time. For example, previous studies have shown that most of the physical properties are usually much less variable over the short distance than chemical properties (Yemefack, 2005). Cost is also one of the factors that lead to minimize the sample size and parameters in many researches. To minimize the redundancy in analysis and get maximum information with a minimum cost, Principal Component Analysis (PCA) and Spearman Correlation Matrix (SCM) are used. Principal component analysis is a data reduction technique that aims to explain most of the variance in the data while reducing the number of variables to a few uncorrelated components (Boruvka et al., 2005; Carroll and Oliver, 2005; Rezaei et al., 2006).

Through ANOVA of soil properties (MSFI) by Soil type (SMU), Land use change (LUCC) and topography of the region, the difference in soil fertility over the 25 years can be explained. MSFI can further be integrated into an indicator measurement of soil fertility (SFI) using a conceptual model of MSFI scoring (adapted from Andrews et al. 2003) and thresholds of soil properties of Rwanda (after Mutwewingabo and Rutunga, 1987).

Soil fertility map of 2006 can then be created from SFI point data and CLORPT grid data using a number of Geostatistical interpolation, and graphical procedures (Mueller et al., 2001) such GLM model. Interpolation using auxiliary variables is most successful for soil attributes whose spatial distribution is strongly influenced by lateral hydrological and slope processes (Park and Vlek, 2002) with respect to soil type.

Through difference computed between SFI 1981 and SFI 2006, the change in soil fertility can be mapped and the extent to which LUCC affect soil fertility can therefore be visualized and explained.

The decision tree (Figure1-2) summarizes the approach used in order to achieve the objective of this research.



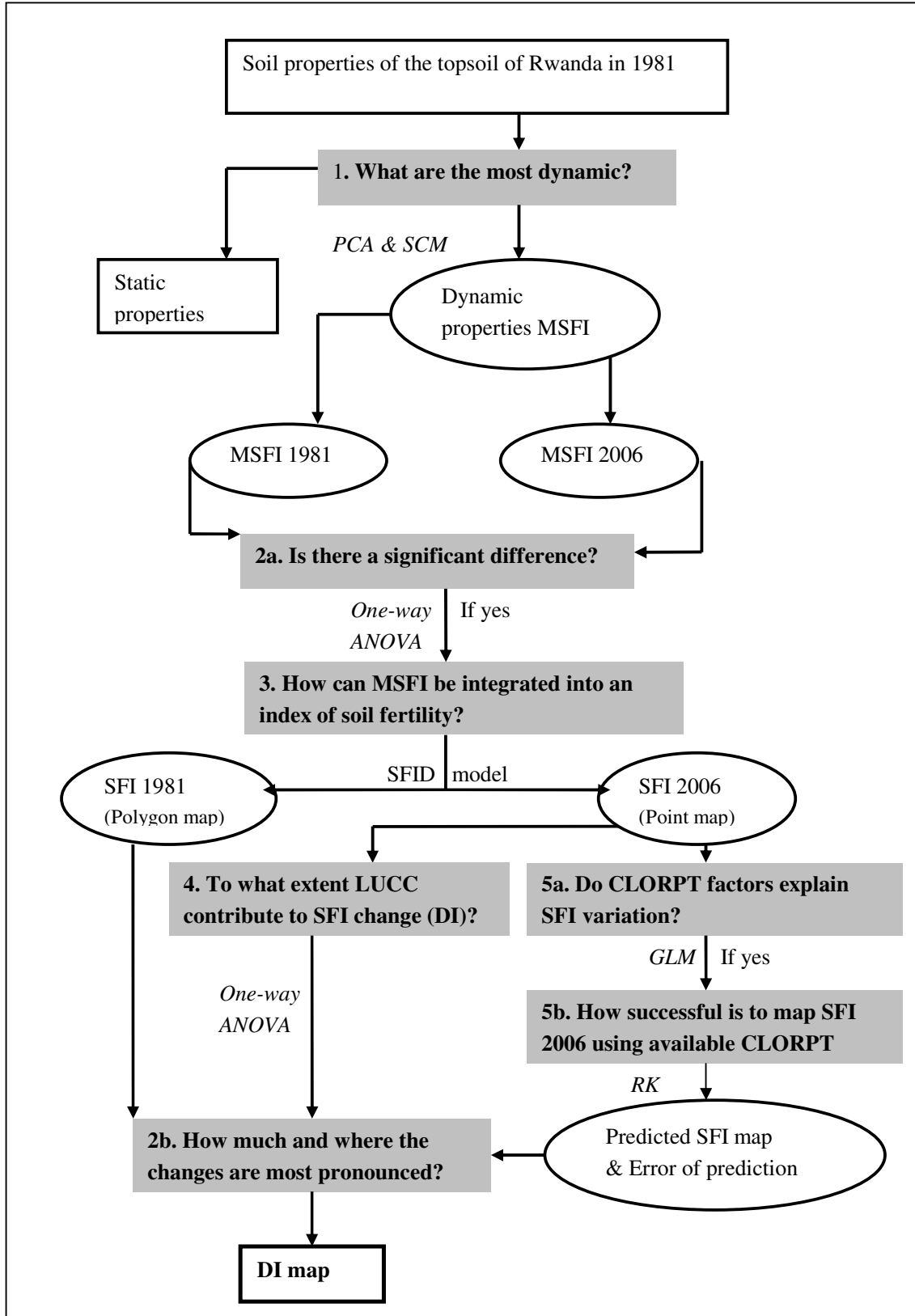


Figure 1-2: Decision tree for mapping and modelling soil fertility degradation using CLORPT approach

## 2. Concepts

### 2.1. Soil fertility degradation in relation to LUCC

Land use and land cover change (LUCC) plays an important role in soil fertility dynamics when compared with natural factors, and can have impact upon soil quality especially under tropical climate conditions. Soil fertility defined as ‘the quality of a soil that enables it to provide nutrients in adequate amounts and proper balance for the specified plants or crops’ has been the cause for much debate and the high fertility theory of tropical soils was dispelled when the forest was cut, crops planted and it was discovered that yield levels were disappointingly low or rapidly declining (Hartemink, 2003). But, this effect of LUCC focuses on short term changes such as deforestation. Other less dramatic decadal scale land use changes turn out to have less effect on soil properties, especially when the results are corrected for landscape variability (Breuer et al., 2006).

An assessment of soil properties upon conversion of natural forests for different purposes is of utmost importance to detect early changes in soil quality. This has been basically proved significant in a tropical forest ecosystem of Bangladesh (Islam and Weil, 2000). Yet, Global change research has stimulated research on the fate of soil organic carbon in relation to soil management and land use change (Pennock and Veldkamp, 2006). However, previous studies have generally not considered landscape relations. These changes might behave differently with regard to lithology variability within the same landscape.

Three different data types are used to assess soil changes caused by agriculture production systems: expert Knowledge, nutrient balances and monitoring of soil chemical properties over time (Type I) or at different sites (Type II). Changes can be assessed by measuring and comparing present values against values at the commencement of the monitoring period (Arshad and Martin, 2002), with historical data when available, with soil attributes under reference ecosystems (Wang and Gong, 1998), or using values measured at different time intervals. This is so called chronosequential sampling or Type (I) data. Type I data show changes in soil chemical property under a particular type of land use change over time whereas with type II data, soil under adjacent different land use systems are sampled at the same time and compared (Hartemink, 2003). Therefore, in this study we focus on both types due to the lower number of type I samples. Moreover, Type II data allows spatial and temporal change while Type I data allows only temporal change analysis.

### 2.2. Integrated approach for soil quality assesment and monitoring

Identification of soil nutrient deficiencies is usually carried out through the analysis of the soil. Nonetheless, this process can be expensive and time consuming, depending on the extent of the area to be evaluated. In other hand, the time series measurements are not always available due to different

reasons. This is the case of monitoring the soil fertility in Rwanda where no soil measurements were taken during the period from 1990 up to now because of the war and genocide 1994 that have been followed by the Rehabilitation priorities. Hence, there is a need of other technology to achieve a reliable monitoring of soil fertility using estimated predictors as an alternative to intensive laboratory measurements that are high costly and time consuming. An integrative assessment framework and the ability of available auxiliary variables such as Digital Elevation data, Soil data and Remote Sensing data to provide explanations of soil fertility change in both future and space without requiring intensive field data collection and laboratory analysis need to be investigated.

During the last decades, mathematical Modelling has become an essential part of ecological research because such models make assessment and predictions in ecological systems more objective and reliable (Jorgensen, 1994). Several study have been done in assessing change of soil properties after deforestation (Lemenih et al., 2005a; Lemenih et al., 2005b) or nutrient depletion on smallholder farming systems (Haileslassie et al., 2005), but few of them integrated this changes into in indicator of land quality.

Indicators of land quality (LQIs) are being developed as a means to better coordinate actions on land related issues, such as land degradation (Dumanski and Pieri, 2000). Economic and social indicators are already in regular use to support decision making at different levels and in some cases for air and water quality, but few such indicators are available to assess, monitor changes in the soil quality. Recently, Andrews et al. (2003) developed the conceptual framework (SMAF) and it has proven to be useful for various soil quality assessment (Andrews et al., 2003). However, He emphasized that the model should be adapted to some situations.

This reason motivates our study to define an integrative index of soil fertility using the thresholds provided for site-specific management. We should now combine probabilistic models provided by mathematicians with previous findings that made available to us, expert knowledge in order to assess the human-induced soil fertility degradation with different auxiliary variables that are already available (Carroll and Oliver, 2005).

While Land refers not just to soil but to the combined resources of terrain, water, soil and biotic resources that provide the basis for land use (Dumanski and Pieri, 2000), soil fertility refers to the critical level of soil properties relative to the requirements of crop production. With respect to predefined classes of soil properties (Mutwewingabo and Rutunga, 1987), an index of soil fertility can then be computed to compare management practices or monitor change over time.

When evaluating the impact of land use change on soils, the soil status under a new land use is compared to a pre-existing steady-state baseline, ideally the native vegetation, which can then be expressed as degradation index (Islam and Weil, 2000; Lemenih et al., 2005a). The deviation of soil fertility from the natural forest gives better indication on how change in land management affects the long-term soil fertility decline. This decline can then be explained by environmental factors such as CLORPT which known as soil forming factor, and see whether the unexplained change can be linked to the change in land uses.

### 2.3. Prediction of soil fertility change using CLORPT model

Global Assessment of Soil Degradation (GLASOD, 1990) has shown that the soil chemical degradation is believed to be important in many parts of the tropics. The loss of nutrient (i.e. soil fertility decline) is severe in Africa (Hartemink, 2003) including a large part of Rwanda.

The method of Geographic information System (GIS) for soil nutrients mapping using different types of interpolation is well proposed and is being used in soil science (Amini et al., 2005; Bishop and McBratney, 2001; Iqbal et al., 2005; Lark and Ferguson, 2004; Li et al., 2004; Liu et al., 2006; McBratney et al., 2003; Mueller and Pierce, 2003; Mueller et al., 2004a; Mueller et al., 2004b; Srivastava and Saxena, 2004). In this chapter, we discuss the various methods that have been used to map the soil properties which are used to monitor the soil fertility change. We also review the soil-environment relationship that has been widely used in soil mapping and modelling.

Jenny's equation (1941) is known as the first model of soil development,

$$S = f(c, o, r, p, t), \quad (1)$$

Where  $S$  represents a soil attribute (e.g. fertility) or soil class,  $c$  (sometimes  $cl$ ) climate,  $o$  organisms including human activity,  $r$  relief,  $p$  parent material and  $t$  time. Most of the soil models are built based on this famous equation. Numerous researchers have taken the quantitative path and have tried to formalize this equation largely through studies of cases where one factor varies and the rest are held constant.

Since the 1960s, there has been an emphasis on what might be called geographic or purely spatial approaches in soil mapping. Soil attribute can be predicted from spatial position largely by interpolating between soil observation locations where soil is considered at some location  $(x, y)$  to depend on the geographic coordinates  $x, y$  and on the soil at neighbouring locations  $(x + u, y + v)$ , i.e.,

$$S(x, y) = f((x, y), s(x + u, y + v)). \quad (2)$$

These purely spatial approaches are almost entirely based on Geostatistics and its precursor trend-surface analysis. Geostatistics provides descriptive tools such as semi-variogram to characterize the spatial pattern of continuous and categorical soil attributes (Amini et al., 2005; Lark and Ferguson, 2004). This technique has been widely applied by soil scientists particularly, various forms of kriging. However, it was recognised early in the development of soil Geostatistics that soil could be better predicted if denser sets of secondary variables correlated with the primary variable were available (McBratney et al., 2003). This technique is called co-kriging, regression kriging, or kriging with external drift:

$$S(x, y) = f(s(x + u, y + v), \{cl, o, r, p, t\}(x, y)). \quad (3)$$

In the early co-kriging studies (e.g. in 1983), these secondary variables were other soil variables, indicating that other soil variables are themselves useful predictors of soil. Later in 1994, with the advent of GIS and improved technology, Odeh et al.(1994) found that co-kriging can be performed with detailed auxiliary data sets of environmental variables derived from digital elevation models and

satellite images (McBratney et al., 2003). In the middle of the 1990s some researchers recognized the similarities between co-Kriging and regression Kriging. In the last approach 'CLORPT' is used to predict the soil property of interest from environmental variables and kriging is used on the residuals. In many cases, kriging combined with regression has proven to be superior to the plain Geostatistical techniques yielding more detailed results and higher accuracy of prediction (Hengl et al., 2004b). Moreover, in several other studies (Bishop and McBratney, 2001; Simbahan et al., 2006), combination of kriging and correlation with auxiliary data outperformed ordinary kriging, co-kriging and plain regression.

Kriging with external drift (KED) is an example of Kriging combined with regression which only allows a linear relationship between the variable of interest and the environmental variables (the external drifts). Although, quantitative relationships have generally been most easily found between soil and topography but the results of many studies illustrate that this empirical relationships between soil properties and terrain attributes are somewhat unique to each soil property and each soil-forming environment. Especially over large areas, predictive capabilities are limited because the relationships between soil properties and landscape attributes are nonlinear or unknown (Lagacherie and Voltz, 2000). In this view, the drift and residuals can also be fitted separately and then summed afterwards (Hengl et al., 2004b). This technique was originally suggested by Odeh et al. (1994, 1995), who named it "Regression Kriging" (RK), whereas Goovaerts uses the term "Kriging after detrending" (Goovaerts, 1999). RK can be more easily combined with stratification, General Additive Modelling (GAM) and regression trees (McBratney et al., 2000). Recently, Multivariate RK with elevation, apparent EC, reflectance, and soil series performed best in terms of increasing map accuracy. For example, in multivariate RK methods relative improvements in map accuracy over OK ranged from 19% to 38% at the three sites in Nebraska and there was little loss of accuracy when sampling intensity was reduced by half (Simbahan et al., 2006).

Therefore, in this study, we focuses on regression kriging (RK) instead of kriging with external drift (KED), not because, RK implies the regression combined with kriging, but it as well allows the nonlinear relationship between the target variable and continuous and categorical predictor. Numerous ancillary variables with potential for soil fertility mapping are available, particularly at landscape scale. Examples include digitized soil surveys (soil mapping units), digital elevation model (DEM) and derived terrain indices. DEM is useful to derive Slope gradient and aspect, the specific Catchment, and the hydrological terrain parameters that are used in Soil loss (Shrestha et al., 2004) as well as soil attributes prediction for a relatively large area (Ziadat, 2005). A methodological approach is also developed (Hengl et al., 2004b). The challenge is to apply them to the local environment, and produce an explicit soil fertility map with regard to covering the variation in primary and secondary variables in feature and geographical space in situation where the sampling intensity is limited.

### 3. Material and Methods

#### 3.1. Gishwati study area in the context of Rwandan environment

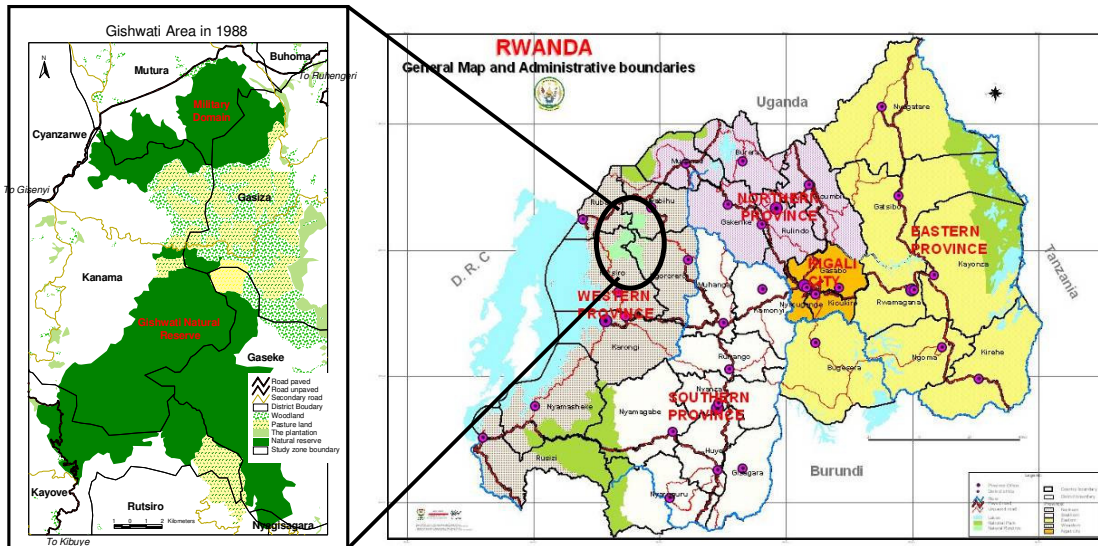


Figure 3-1: Map showing Gishwati study area before 1995

##### 3.1.1. Climate, geology and geomorphology

Rwanda occupies the eastern shoulder of the Kivu–Tanganyika rift in Africa. It lies between latitudes 1°04' and 2°51' South, and between longitudes 28°53' and 30°53' East. The general altitude increases from about 1000 m at the edge of the Lake Victoria basin in the east to over 2600 m at the crest of the rift shoulder in the west. The lowest point in Rwanda goes down to 950 m, the highest reaches 4507 m. Due to its elevation, Rwanda enjoys a rather mild climate for a country so close to the equator. The mean annual temperature turns slightly around 20°C. Annual rainfall varies between 700 and 1,400 mm in the eastern and western lowlands, between 1,200 and 1,400 mm in the Central Plateau and between 1,400 and 2,000 mm in the high altitude region (MINITERRE/Rwanda, 2003). The two rainy seasons extend from the middle of September to December and from the end of January till May or June, respectively. The annual distribution of the orographic rainfall, issued from the Indian Ocean during the passage of the intertropical convergence belt, reflects the general topography (Moeyersons, 2003). According to Köppen classification system, most of high elevation regions including Gishwati study area are belong to the CW-type with 2-3 months of rainfall under 60 mm (ABOS-AGCD, 1983).

The topography forms of Rwanda are the result of the combined action of erosion and of tectonic movements. After the metamorphosis and folding of sedimentary rocks that gives rise to of Rusizian

and Burundian, the northern part of the country, including the Gishwati study area is underlain by Precambrian ruzizian rocks composed of a wide range of shales locally pierced by granitic batholiths. These granites give way to more rounded hills and valleys than the shales and result a “*landscape of thousand hills*” with steep slope that characterized the “*Congo-Nile watershed divide*” in West (Moeyersons, 2003).

The investigated area is within the Congo-Nile watershed divide in Western province of Rwanda (Figure 3). This area was chosen because of its representative of complex lithology and landscape diversity due to elevation differences from valley floor to mountain summits, and related land use changes having influence on soil erosion which is considered typical for the highland of Rwanda. It extends from easting of 29°21'40” to 29°28'50” longitude and southing of 1°36'52” to 1°52'17” latitude.

### **3.1.2. Soils and fertility**

Soils of Rwanda present a high variability in physical and chemical properties. As reported by different researchers, most soils are fine textured, but the soil depth is strongly variable as well as weathering intensity and chemical soil fertility (Mukashema, 2003; Mutwewingabo, 1984; Neel et al., 1976; Ntaneza, 1988; Zaag, 1981; Zaag et al., 1982). Consequently, most of the Soil Taxonomy (Soil Survey Staff, 1983) orders are found in Rwanda (Verdoodt and Van Ranst, 2006), except Spodosols (ABOS-AGCD, 1983).

In high elevation region where Gishwati watershed is located, Entisols occupy the river valleys; these are mostly Aquents. Histosols are very common in the poorly drained swamps. They are not typical for Gishwati but they occur elsewhere in Rwanda where the drainage is low. In the zones covered by volcanic material and, elsewhere on steep slopes, the Inceptisols are an extensive soil unit. Many soils developed in the colluviums accumulated at the foot of the hill slopes belong to this order too. Most well-drained soils of this region are belonging to Ultisols. In spite of the steep slopes, soils are generally deep and rich in organic matter. Some of the soils show a structure which could make them considered as Oxisols but generally their exchange capacity exceeds the value which is necessary for Oxisols (ABOS-AGCD, 1983). Speaking about soil fertility, we must stress that besides the classic pedogenetic factors, human activity did largely influence to the soil fertility status. This human influence, together with relief, explain why there is a wide range in crop yields over small distance, even in soils developed in the same parent material.

### **3.1.3. Historical land Use and Land cover change**

The change in land use in Rwanda especially in Gishwati study area is result of the changes in population, which doubled nationally between 1978 and 2002. Over 90 percent of the population relies on subsistence agriculture to meet its needs, with a concomitant need for land, which puts great pressure on the country’s remaining natural ecosystems, whether forested, savannas, or wetland (Plumptre et al., 2001).

Since 1980s, Gishwati forest reserve had been heavily affected by human activities prior to the Rwandan civil war. It constituted approximately 280 square kilometres in the mid-1970s and contained populations of chimpanzees (*Pan Troglodytes*) and golden monkeys (*Cercopithecus mitis kandti*), although the forest was fairly degraded by many years of cattle herding within the forest. The World Bank supported an integrated forestry and livestock project that converted 100 square kilometres to pasture and another 100 square kilometres to pine plantations in the early 1980s. A 30 square kilometres area was designated as a military zone in the north of the forest, leaving only 50 square kilometres of natural forest.

During and following the war in 1994, the northern part of Gishwati was used for camps of displaced persons, which grew rapidly. People settled and farmed within the reserve, thus creating further pressures on land and deforestation. In early 2000, the Nyungwe forest conservation project (PCFN), supported by the Wildlife Conservation Society (WCS), organized a survey of Gishwati natural forest to assess the current status and to determine whether it would be useful to encourage conservation efforts. There was little of the original forest remaining in Gishwati. Only a few stands of trees of less than one hectare in size within cropland were observed (Plumptre et al., 2001).



### 3.2. Research Methods

#### 3.2.1. Methodological flowchart

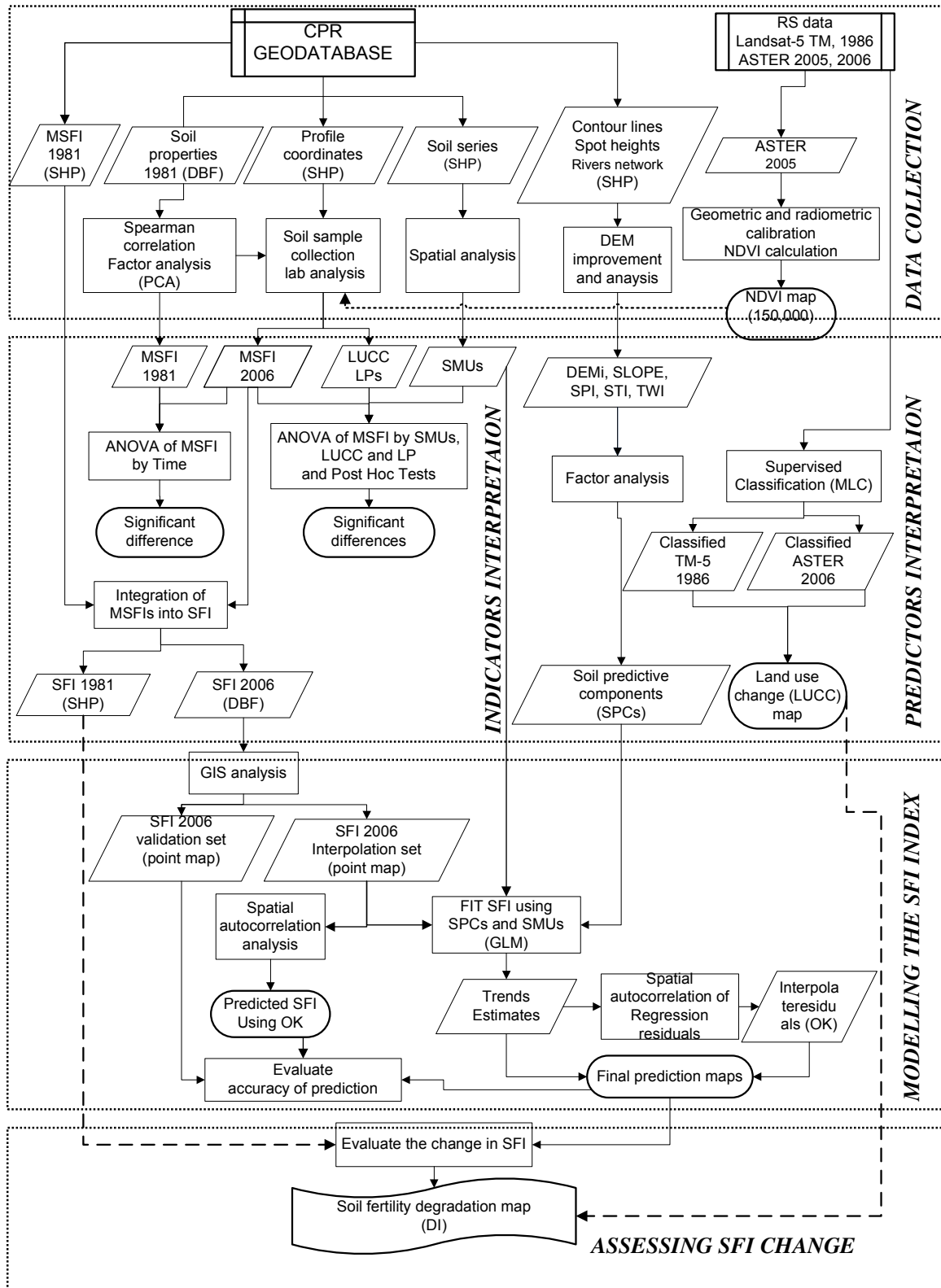


Figure 3-2: Methodological flowchart soil fertility change analysis and mapping.

### 3.2.2. Spatial and temporal boundaries of the study

To assess soil fertility decline, it is necessary to define the spatial and temporal boundaries of the system under study. The loss of fertility by erosion at catchment scale is measured spatially using the **black box** approach. This approach considers the depth, the width and length as important boundaries of the box. The same approach is used to explain the transfer of nutrients from one area or spatial scale to another by subsurface flow (Hartemink, 2003). This approach was applied to this study. Gishwati catchment represents a catena of about 280 km<sup>2</sup> with flows of nutrients from upper slope to valley by erosion or subsurface flow process. The depth of the box was about 30 cm as the most vulnerable layer to erosion and most important for crop growth. The study focused firstly on different land uses for the entire area and for the temporal study, the profiles of 1981 project were also revisited.

### 3.2.3. Data types

#### 3.2.3.1. Soil fertility indicators

To assess the change in soil fertility at a given site, Type I data were used. This method is also called chronosequential sampling or Type (I) data. Type I data show changes in soil chemical property under a particular type of land use change over time. Usually the original level is taken as the reference level to investigate the trend in such changes. Example, Type I data have been used for quantifying soil contamination by comparing soil samples collected before the intensive industrialization period with recent samples taken from the same location (Hartemink, 2003). The same approach was used to quantify the change in soil fertility of the study area. We used the chemical properties already measured from 1981 through the semi-detailed soil survey done during the project named “*Carte pedologique du Rwanda*” (CPR), and compared to newly collected and analysed soil samples of 2006.

Because there were only 17 points of Type I which was not even distributed in different land uses, the land use change effect on soil fertility was assessed using Type II data. In this approach, soil fertility status under adjacent different land use systems were sampled and compared. This is so called biosequential sampling (Hartemink, 2003) or synchronic sampling (Yemefack, 2005). The main underlying assumption is that the soils on the cultivated land, pasture, planted forest and reference land use (in this case natural forest) are the same soil type (refer to great group units), but that the differences in soil fertility can be attributed to the difference to the differences in land use. Revisiting the site is the key for Type I data, while knowing the historical land use is much more important for Type II. Both types were involved in assessing the spatio-temporal change of soil fertility across Gishwati catchment area. Type II was useful for spatial distribution of the change whereas Type I was important for temporal change analysis.

### 3.2.3.2. Site specific explanatory variables

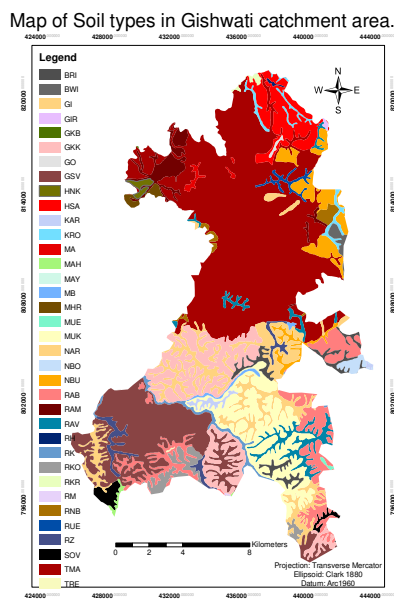
Site-specific information on historical land use change and related management and landscape position were recorded during soil sample collection (Appendix 8-1). GPS 12 XL was used to record the geographical position of each sample site.

### 3.2.3.3. Soil fertility predictors

#### a) Soil data

A digital soil map (1:50000) of the study area were obtained from the National Soil Geodatabase in the Ministry of Agriculture (Table 3-2). The soil map resulted from the soil survey of Rwanda which started in 1981 and finalized in 1994. Initially, the intention of the soil survey was to map Rwanda at scale 1:100,000. However, the geologic and geomorphologic complexity of Rwanda and the multiplication of rural projects required more detailed soil information, which resulted in a modification of mapping scale to 1:50,000. This semi-detailed soil survey, based on extensive use of aerial photographs and fieldwork, was accomplished. From 1989 onwards, the soil maps and all observation points with their corresponding data were stored in a master database using GIS and relational database software (Verdoodt and Van Ranst, 2006). Stopped in 1994 due to the war, the digital storage of the soil data was later finalized at Gent University, Belgium (1998-2000). Both the activities in Rwanda and at Ghent University were financed by BADC (Belgian Administration for Development Cooperation).

**Figure 3-3: Soil series map of Gishwati study area**



The national survey resulted in the elaboration of 43 soil maps, at a scale 1:50,000, covering the whole of Rwanda. More than 2000 soil profiles, corresponding to 176 different soil series had been described and analysed among them 36 series are found in the study area (Figure 3-3, Appendix 2 for series' description). Gishwati watershed is located in two soil maps (GISENYI and MULUNDA). The Automation of the data started with the digitizing of hardcopy maps by use of the GIS software ARC/INFO. Each soil unit received a unique label that was related to a numerical database with the tabulated properties of each soil series.

The spreadsheets containing the profile description and analytical information were imported in Access database. Relationships were built between three tables containing the general profile information, the horizon description and the horizon analytical data. Through use of unique soil

profile number, these numerical data can be easily linked to cartographic data (Verdoodt, 2003). About 43 profiles are located in Gishwati watershed but only 17 profiles among them are within the study area.

The acquired base information on soils was then processed for the purpose of this study. Soil map of Gishwati catchment area was 36 soil mapping units (refer to the series in Soil Taxonomy). Due to their similarity in soil properties those were reclassified and merged in 14 great groups (appendix 8-4) using ArcGIS 9.1 software. Soil samples were distributed in 8 major great groups of the study area. Those great groups were considered in soil fertility change interpretation. Soil properties of 1300 profiles at horizon A were extracted from the spreadsheet of soil properties of Rwanda which contain more than 7000 records of soil variables for all soil profiles and all horizons of each profile. Those were used in indicator section procedure.

#### b) Vegetation cover data

Prior to the field data collection, a proper visualisation of Land use and cover is important to guide on the criteria of selecting sample design and distribution of sample points across the study area. NDVI map was prepared for this purpose.

Aster image level-1B (L1B) of February 21<sup>st</sup> 2005 was acquired (Table 3-2) and processed using ILWIS scripts. Visible and near infrared bands (VNIR) were imported and reprojected to local projection (Transverse Mercator), Ellipsoid (Clarke 1880), and Datum (Arc 1960) to prevent the shift and match with other thematic layers). Then, the image was georeferenced to the corner coordinates of the study area. ASTER Level-1B data are offered in terms of scaled radiance as sensor calibrated Digital Numbers (DN). To convert DN to radiance at the sensor, the unit conversion coefficients were used. Radiance (spectral radiance) is expressed in unit of  $Wm^{-2} sr^{-1} \mu m^{-1}$ . The relation between DN values and radiances is expressed in equation 4. The maximum radiances depend on both the spectral band and gain setting. The sensor calibrated DN values are converted to spectral radiance in ILWIS using the unit conversion coefficient of each band as follows:

$$L = (DN - 1) \times C \quad (4)$$

Where L is calculated spectral radiance ( $Wm^{-2} sr^{-1} \mu m^{-1}$ ), DN is the value of sensor calibrated digital number and C is the unit conversion coefficient from the metadata in HDF file. Unit conversion coefficient used for different bands and for different gain settings are shown in table 3-2.

The image was corrected for artifacts due to atmospheric conditions by computing the exoatmospheric reflectance of each image pixel using equation 5. Reflectance ( $\rho$ ) is defined as the wavelength dependant ratio between reflected and incoming energy and can be expressed as:

$$\rho = \frac{L_{\lambda} \pi d^2}{ESUN_{\lambda} \cos \theta_z} \quad (5)$$

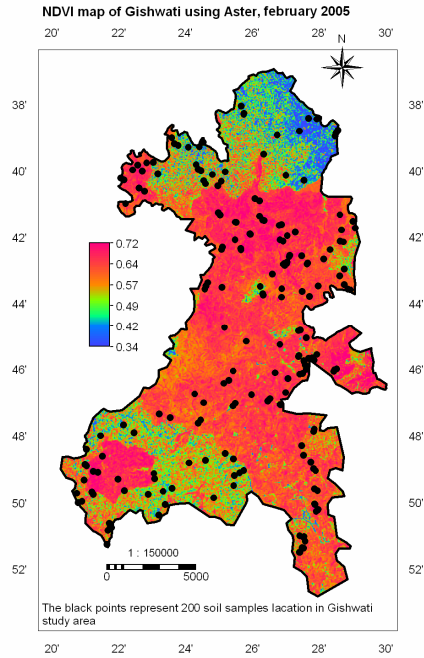
Where  $\rho$  represents the top of atmosphere reflectance,  $L_{\lambda}$  Radiance at the sensor ( $Wm^{-2}sr^{-1}\mu m^{-1}$ ),  $d^2$  Earth Sun distance (AU),  $\theta_z$  solar zenith angle (degrees) and  $ESUN_{\lambda}$  the mean exoatmospheric irradiance ( $Wm^{-2}\mu m^{-1}$ ). Note that earth sun distance depends on the day of the year (Julian day) and the solar zenith angle depends on day and time of acquisition of the image, the latitude and the longitude of the location. Earth Sun distance is given in table 3-2 (Aster user handbook).

NDVI map as ratio index of vegetation cover was then calculated to guide field data collection, using the following formula (Equation 6):

$$(\rho_{NIR} - \rho_R) / (\rho_{NIR} + \rho_R) \tag{6}$$

Where  $\rho_{NIR}$  is the reflectance on near infrared band and  $\rho_R$  is the reflectance on red band. NDVI map enabled us to distinguish different entities of land use in the study area (Figure 3-3). Thus, were followed in data collection.

**Figure 3-4: NDVI map of Gishwati study area using Aster, February 2005**



**Table 3-1: Unit conversion coefficients and mean exoatmospheric irradiance of ASTER bands**

Bands	C	ESUN
1	0.676	1846
2	0.708	1555
3N	0.862	1120
4	0.217	231

**c) Land use and land Cover change data**

To map change in land use, Landsat-5 TM image of July 19<sup>th</sup> 1986 and ASTER image of June 16<sup>th</sup> 2006 were acquired and processed to a common 15 X 15 m grid by resampling method. They were reprojected to the local projection (refer to the previous paragraph). Supervised classification using Maximum Likelihood classifier (MLC) was performed in ERDAS Imagine 8.7

package for both images. This method was found to be the most accurate and objective when the results were compared with the changes recorded in the CPR database during soil survey 1983, and information recorded during soil sample correction, September 2006. With the MLC, all significant land cover types in the image were designated as classes. The overall classification accuracy was assessed by comparison of 43 cover points recorded by CPR project for Landsat-5 TM, 1986 and 200 sample points recorded during field work for ASTER, 2006. The classified images for each date were combined into one file and change images were created representing pixels that changed from Natural forest to pasture in 1980s, from Natural forest to forest plantation (*Pinus* and *Acacia sp*), and those that changed recently from natural forest to cropland after 1995.

Change map was then confronted to the change in soil fertility in order to analyze the extent of human- induced soil fertility change in Gishwati watershed.

**d) Topographic data**

Terrain attributes derived from digital elevation models (DEM) are commonly useful explanatory variables in predictive soil models (Sonneveld et al., 2006; Tomer and James, 2004; Ziadat, 2005). The contour data of 25 m equidistance were acquired to generate different terrain parameters. This contour map was created in the same project as soil map. During the CPR project, 43 topographic maps at scale 1:50,000 were produced by 1987. This map contains contour lines at an equidistance of 25 m. Digitization of the topographic map data was realized by scanning the hardcopy maps and vectorising, georeferencing and geocoding of the digital data. Arc view Software and 3-D Analyst

extension were used to derive a digital terrain model (DTM) for each map sheet (Verdoodt, 2003). The river network and spot heights of the study area were also supplied in the same Geo-database. Prior to the calculation of terrain parameters, the quality DEM was improved using the method proposed by Hengl et al., 2004 (Hengl et al., 2004a). This procedure is done to account for the features that are not shown by the contours such as ridges and valley bottom. The spot heights were assigned to the medial axis between the closed contours and the river networks were used to adjust the final DEM. The sinks were also filled. All these steps are well explained in terrain analysis user guide (Hengl et al., 2003). The improved DEM was then used to generate the relief parameters for soil landscape modelling.

The contour was rasterized using a common grid (15x15m) of all variables involved in this study. This was used to generate Digital Elevation Map (DEM) and terrain attributes such as slope gradient ( $S = \tan \beta$ , percent), specific catchment area ( $A_s$ ,  $m^2 m^{-1}$ ), topographic wetness index (TWI), stream power index (SPI) and Sediment transport index (STI).

The topographic wetness index, a predictor of zones of soil saturation, is the ratio of specific catchment area to slope gradient:

$$TWI = \ln(A_s / S). \quad (7)$$

The stream power index, a measure of runoff erosivity, is the product of specific catchment area and slope gradient:

$$SPI = \ln(A_s \times S). \quad (8)$$

The sediment transport index, also called erosion index is modelled as following:

$$STI = (AS / 22.13)^{0.6} \times (\sin \beta / 0.0896)^{1.3}. \quad (9)$$

For detail elaborations refer to (Hengl et al., 2003; Thompson et al., 2006; Tomer and James, 2004).

The calculations were performed using flow indices script in ILWIS 3.0.

**Table 3-2: Data type and key variables considered for soil fertility change analysis**

Data type	variables	Source
Initial Soil fertility indicators (before deforestation)(1:50,000)	pHw, pHKCl avail.P, OC, Exch. bases (Ca, Mg, K), Echange acidity (Al, H).	Soil Geo-database of Rwanda MINAGRI (Kigali/Rwanda)
Actual MSFI (2006)	pHw, pHKCl, avail.P, OC, Exch. bases (Ca, Mg, K), Exch. acidity (Al, H).	Primary data Soil measurements
Soil type	Soil series and related variables	Soil Geo-database of Rwanda MINAGRI (Kigali/Rwanda)
Topographic data (1:50,000)	Contour data (25 m equidistance) Spot heights and River network DEM, derived Terrains parameters	Soil Geo-database of Rwanda MINAGRI (Kigali/ Rwanda)
Interviews and Field observation	Historical land uses and related management Erosion features	Site-specific information (smallholder farmers and local operators: ISAR/Gishwati, PAFOR).
Remote Sensing data Landsat-5 TM 16 <sup>th</sup> July 1986, ASTER 21 <sup>st</sup> January, 2005 and 21 <sup>st</sup> June 2006	Land use / cover types Vegetation index (NDVI)	Geodata Warehouse (ITC Enschede)

### **3.2.4. Soil fertility change analysis**

#### **3.2.4.1. Selection of the Minimum soil fertility indicators (MSFI)**

Due to the limited logistic and time, the study of soil fertility as the combination of various soil properties that determine the capacity of the soil to crop production function, started by reducing the soil variables prior to data collection and laboratory activities. This led to a minimum data set for soil fertility assessment in Gishwati catchment area. The MSFI should just be considered as the smallest set of the soil chemical properties that can best represent the human-induced change in soil fertility. This approach is similar to the Minimum Data Set (MDS) approach (Park and Vlek, 2002; Yemefack et al., 2006), the only difference is that the MSFI is limited to the chemical soil properties whereas the MDS consider both physical and chemical properties i.e. the MSFI is part of the MDS.

We agree with different researchers that the soil chemical properties to be included in a MSFI must be sensitive to changes in soil management, soil perturbations, and inputs into the soil system. (McBratney et al., 2003; Sena et al., 2002; Yemefack, 2005; Yemefack et al., 2006; Yemefack et al., 2005). Each selected property must also be inexpensively, easily and reproducibly measurable.

Reducing the redundancy between soil chemical variables was achieved by evaluating the correlation of variables over the whole dataset of Rwanda. The Spearman correlation was computed on the multivariate data matrix using 1300 soil samples collected and analyzed during the semi-detailed soil survey of 1981. This non-parametric method was used to avoid distortions from non-normally distributed variables or extreme values (Yemefack, 2005). To cross-check the result of this correlation analysis, Principal Component Analysis (PCA) was also performed as one of factor analysis for soil variables reductions. Properties that had high score of PCA value and highly correlated were classified in one group. The included soil variable in the MSFI was defined to be the highest scored variable in PCA and sometimes times with less correlation among themselves.

#### **3.2.4.2. Sampling design**

Refer to the distribution of human activities in Gishwati study area, such as the division of the land into different land uses (Figure 3-3), we realised that there were organized control. As result, the human-induced soil fertility change can not be random, except within the same use, same soil and same landscape position. This situation leads us to the stratified sampling design. We agree also with soil conservation scientist that simulated erosion patterns and nutrient losses are directly related to the flow network of the catchment. Since the model routes water and suspended sediment towards the outlet using a user-supplied network (Jetten et al., 2003), the flow of soil nutrients is closely related to this network. Therefore, stratified toposequence transect sampling seemed to be appropriate and transect location was chosen purposely. About 67 toposequential transect locations were chosen to represent major Land Use classes with different topography and soil types. Each transect included at least three of five separate sampling points collected at summit, upper slope, middle slope, lower slope, and valley.

### **3.2.4.3. Soil sample collection and laboratory analysis**

Nutrients in soil solution are readily plant-available. In that case, topsoil properties may be used as an indication of nutrient availability to plants because most roots are concentrated in the A horizons (Lilienfein et al., 2003). Soil samples were collected from Gishwati catchment area by augering at 0-30 cm depth. These were analysed in the PASI laboratory, Faculty of Agriculture at the National University of Rwanda (NUR) for the following determinations: pH water and pHKCl with a ratio of 1:2.5 (pHw), organic carbon (Org.C) using the Walkley-Black method, available P (avail.P) using the Bray-1 method, exchangeable bases (exch.B) using the ammonium acetate percolation method, exchange acidity (Exch. Ac.) using an unbuffered KCl solution. All these methods are described in (ABOS-AGCD, 1983). The same methods were used to analyze the soil chemical properties in CPR project.

### **3.2.4.4. Relations of soil fertility to predictive factors**

One-way analyses of variance (ANOVAs) were performed on each soil fertility indicator per Soil type, Type of change in land use and landscape positions to test whether, the relations investigated were statistically significant. Mean values were compared using Duncan test. Duncan test separates the means of soil fertility indicator variables at  $\alpha = 0.05$ .

## **3.2.5. Soil fertility change interpretation**

### **3.2.5.1. Development of soil fertility index (SFI)**

The development of an integrative index of soil fertility (SFI) was utmost important to capture the variability of individual indicator of soil fertility. Once we had the most dynamic soil variables, the next task was to integrate them into a value that indicates at once the maximum variability both in space and time. This index gives an explicit indication of soil fertility that can not be easily seen using each soil property.

Probabilistic model are becoming increasingly important in analyzing the huge amount of data being produced by different scale, methods or different laboratory analysis. The integration involved transformation of each observed MSFI value in scored value (probability value) using thresholds for interpretation of topsoil properties of Rwanda (Appendix 11, 12, 13). This method assumes that the indicator is measured according to the standard method for near surface (0-30 cm) (Mutwewingabo et Rutunga, 1987), and that sampling design was appropriate for the area to be assessed (Andrews et al., 2004).

The conceptual model (Figure 3-5) summarizes the steps for soil fertility index development (SFID). In this framework, measured MSFI values were transformed into unitless score (0 to 1) using thresholds of soil properties classes of Rwanda and probabilistic approach. Each soil fertility indicator was assigned its probability that it falls into very high fertile soil. An indicator score of 1 represent



the highest potential function for the production system. That means the indicator is nonlimiting to soil productivity function.

The developed SFI index was defined as the probability that any type of soil with its measured soil properties (MSFI values in this case) falls into good fertile soil. SFI vary from 0 to 1 which means from extremely low fertile soil to very high fertile soil (Equation 10).

$$0 \leq SFI \leq 1 \tag{10}$$

Each MSFI was assigned a score equivalent to its probability of falling in very high fertile soil (i.e. SFI=1) by using the threshold and soil property classes developed by Mutwewingabo et al. (1987). The classes (c) must be mutually exclusives according to the axiom of the probability in general. The number of classes leads to the probability assigned to each class as expressed in equation 11.

$$p_c = \frac{1}{n_c} \tag{11}$$

Where,  $p_c$  represents the probability the class c, and  $n_c$  the number of classes. The score ( $Sc_i$ ) given to each soil class depend on its position in class' range and  $p_c$  as in equation 12:

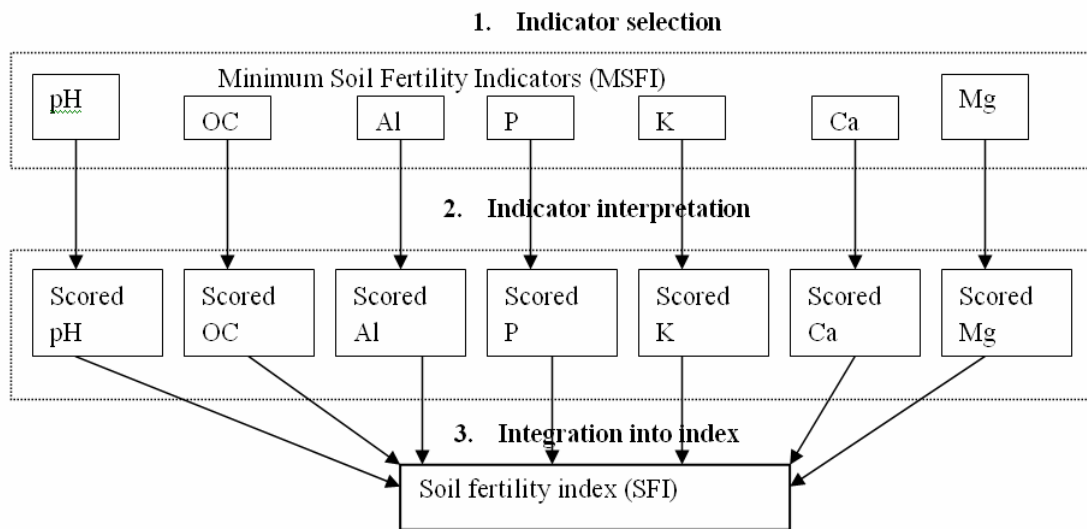
$$Sc_i = c_j \times p_c \tag{12}$$

Where,  $c_j$  represents the class number which varies from 1 to j depending on the amount of classes for the MSFI interpretation (refer to appendices 11, 12 and 13). Note that for all MSFI, "More is better" except for Al where "Less is better".

The scored indicators were combined to form a single index value reflecting soil fertility index (SFI) of each sample site. This index value is considered to be an overall assessment of soil quality of a given site. SFI is an additive index and it is accomplished by summing the scored value for each soil fertility indicator, dividing by the total number of indicators and then multiplying by 10 (Eq.10):

$$SFI = \left( \frac{\sum_{i=1}^n Sci}{N} \right) \times 10 \tag{13}$$

Where  $Sci$  represents the scored MSFI indicator value and N is the number of indicators of MSFI. The data was multiplying by 10 to provide index values in range (1 to 10 rather than 0 to 1). Andrews et al, 2004 found to be more amenable for producers and others potential users.



**Figure 3-5: Conceptual model of soil fertility index (SFI) development after Andrews et al. (Karlen et al., 2003)**

### 3.2.5.2. Index of Soil fertility deterioration (DI)

After that SFI was determined for each sample point, the soil deterioration index (DI) was computed on the assumption that the status of soil fertility under pine plantation, pasture and agriculture were once the same as that of adjacent soil under natural forest prior to conversion. The difference between each value of SFI under pine plantation, pasture and agriculture were compared to baseline mean value of SFI under natural forest. The deterioration index (DI) was calculated as deviation of SFI values from SFI of Natural forest ( $SFI_{NF}$ ) to other land cover type ( $SFI_{LC_i}$ ) under a specific soil type. Eutrandspts were used as one of representative soil type in Gishwati study area.

$$DI = SFI_{LC_i} - SFI_{NF} \quad (14)$$

### 3.2.5.3. Geostatistical analysis

The development of the cross variogram model for soil fertility might be very beneficial for Gishwati study area if prediction is sufficiently accurate. For instance in Agriculture, the predicted map from developed SFI model might be a tool for monitoring and implementing site specific management. Secondly, in management goal, SFI define at once the situation that happened instead of what happened to individual parameter which could be difficult to understand because sometimes they contradict each other. In this case SFI average the behaviour of its predictors into a common behaviour of soil fertility across the study area.

#### 1) Spatial prediction of SFI using smoothing Kriging (OK)

Kriging is one of interpolation techniques that use spatial correlation between observation points. With OK, the trend is modelled as a function of the location (Hengl et al., 2004b). There are a number of considerations in geostatistical analysis including normality, anisotropy, and semivariogram model selection. While normality is not a requirement for developing the semivariogram or kriging, the classical linear Kriging predictor (i. e. OK in this case) is not the best predictor if the spatial process is not Gaussian (Mueller et al., 2001).

The interpolation of target variable (in this case SFI) is based on assumption that the original process is a diffusion process (Lark and Ferguson, 2004; Webster, 2000). It varies continuously so that if the target variable takes values  $z_j$  and  $z_i$  at locations  $x_i$  and  $x_j$ , respectively, then all intervening values between these two must occur at locations between  $x_i$  and  $x_j$ . The commonest model, which we used for Kriging, is the Gaussian diffusion process. Since SFI data more or less resembled to Gaussian distribution, the nonlinear transform to achieve normality was not necessary. A variogram is estimated using *Matheron's estimator* (1965).

$$\hat{\gamma}(h) = \frac{1}{2M_h} \sum_{i=1}^{M_h} \{z(x_i) - z(x_i + h)\}^2 \quad (15)$$

Where,  $Z(x_i)$  is the property value (SFI in this case) at a given location ( $x_i$ ),  $h$  is lag (both distance and direction),  $M_h$  is the pairs of observation points, separated by  $h$ ; and  $\hat{\gamma}(h)$  is semi-variance at lag  $h$ . Kriging assumes a certain degree of spatial correlation between the input point values. To investigate whether SFI values were spatially correlated and until which distance from any point this correlation occurs, we used the Spatial Correlation operation in ILWIS. This distance value was then used for the limiting distance in point interpolations using OK.

## 2) Spatial prediction using Kriging with ancillary variables

SFI pair points may be correlated because they are affected by similar processes, or phenomena, that extend over a larger area. For instance they may correlate because they are under similar land use or under the same soil type.

In this case, a mixed approach is used to explain what can be explained by knowledge of soil-forming factors (as expressed in CLORPT model), and then see if the remaining unexplained variability has any geostatistical relation which can be used to improve the prediction. Regression Kriging (RK) or Kriging with External Drift (KED) use a CLORPT variable as predictor of soil attribute. However, both KED and RK has the same mathematical form and yields the same predictions (Hengl et al., 2004b). The impact of soil type, land use change and landscape on soil fertility variation in the study area were investigated using analysis of variance procedures. Most of the soil fertility variables varied between soil types, type of change in land uses and landscape positions. Therefore, the interpolation involved them in the model of SFI trend by using stepwise regression procedures and the residuals were extracted and fitted in geostatistical model.

We used DEM, Slope gradient (S), Topographic Wetness Index (TWI), Stream Power Index (SPI) and Sediment Transport Index (STI) as predictors. They were first linearly stretched to a range of 0-255 to give each map equal contrast. To account for multicollinearity, a factor analysis prior to regression analysis was applied to produce the soil predictive components (SPCs) using ILWIS. The use of standardised principal component instead of the original terrain variable improves the prediction for soil-landscape modelling (Hengl et al., 2004b). The SPCs values were then extracted for SFI locations for the stepwise regression modelling. The regression model of SFI as function of SPCs was then generated.

SFI Point data were imported to an ILWIS table with XY and SFI coordinates. We first estimated the values of predictors (SPCs) at the measured SFI locations and we fitted the trend (SFI trend) by using an obtained regression model. Secondary, we estimated the residuals at sampled locations using: SFI residual as difference between measured SFI and SFI trend using 'SFI-MapValue (SFI trend, Coordinate)' syntax. Third, Gaussian variogram model of SFI residual was fitted and SFI residual was interpolated using OK (interpolated SFI residual). The fitted trend and residuals was then added back together using 'predicted SFI=SFI trend +interpolated SFI residual'. All those calculation was done using ILWIS command line (for detail refer to <http://spatial-analyst.net/RKguide.php>).

To improve the SFI prediction, SMUs was used to stratify SPCs in Stepwise regression by using WLS. This technique adds the contribution of soil type on SFI to SPCs explanation. Then RK proceeded in ILWIS as described in previous paragraph.

## 3) Spatial prediction of soil fertility deterioration over 25 years

The spatial distribution of soil fertility deterioration index (DI) was computed as deviation of SFI from SFI before deforestation to the current situation of soil fertility. In this case Type I method is made possible through the predicted maps because it gives on opportunity to monitor the change in soil fertility at a given location. This deterioration reflects the human-induced soil fertility loss in Gishwati study area and it is expressed as in equation 16:

$$DI_{(x,y)t} = f(SFI_{(x,y),t}) = SFI_{(x,y)t_f} - SFI_{(x,y)t_i} \quad (16)$$

Where, DI represents the current deterioration index at  $x, y$  location,  $SFI_{(s,y)t_i}$  the soil fertility index at time reference ( $t_i$ ) and  $SFI_{(s,y)t_f}$  the soil fertility at current time ( $t_f$ ).

#### 4) Model quality evaluation

The evaluation of soil fertility prediction map was done using interpolation and validation sets. From 200 soil observations (N), 30 observations were randomly selected as validation set ( $n$ ). The true prediction accuracy was then evaluated by comparing estimated values of soil fertility ( $z^\wedge(SFIj)$ ) with actual observation at validation points  $z^*(SFIj)$  in order to assess a systematic error, calculated as mean prediction error (MPE) using Equation (17).

$$MPE = \frac{1}{n} \cdot \sum_{j=1}^1 [z^\wedge(SFIj) - z^*(SFIj)] \quad (17)$$

And accuracy of prediction, calculated as root mean square prediction error (RMSPE):

$$RMSPE = \sqrt{\frac{1}{n} \cdot \sum_{j=1}^1 [z^\wedge(SFIj) - z^*(SFIj)]^2} \quad (18)$$

In order to compare different methods of SFI prediction, the error was then normalized by the total variance of observed samples. This is so called the normalized mean square error (NMSE) or relative prediction error (Park and Vlek, 2002):

$$NMSE = \frac{\frac{1}{n} \cdot \sum_{j=1}^1 [z^\wedge(SFIj) - z^*(SFIj)]^2}{S^2} \quad (19)$$

Where,  $S^2$  is the total variance of SFI index at observed sample. As a rule of thumb (Hengl et al., 2004b), we considered that a value of relative prediction error (NMSE) close to 40 % means a fairly satisfactory accuracy of prediction. Otherwise, if the value exceeds 71%, this means that the model accounted for less than 50 % of variability at the validation points and the prediction is unsatisfactory. The goodness of fit was also confirmed by adjusted coefficient of determination  $R_a^2$  and Pearson's correlation coefficient (R). Note that  $R_a^2 \geq 0.85$  correspond in many cases to the relative prediction error of  $NMSE \leq 0.40$  (Hengl et al., 2004b; Park and Vlek, 2002).

The uncertainty of predicted SFI map was visualized by using predicted error map. The error map was computed as the standard error or standard deviation ( $\sigma$ ) of the predicted SFI from measured SFI point location (equation 20).

$$\sigma = \sqrt{(\sum_i (W_i \times \gamma(h_{\rho_i})) + \lambda)} \quad (20)$$

Where,  $\sigma$  is the standard error or the standard deviation of the output pixel estimate,  $h_{\rho_i}$  is the distance between the output pixel  $\rho$  (i.e. predicted SFI) and input point  $i$  (i.e. measured SFI),  $\gamma$  is the value of the semi-variogram model for the distance  $h_{\rho_i}$ , i.e. the semi-variogram value for the distance between the output pixel  $\rho$  and input point  $i$ ,  $W_i$  is a weight factor for input point ( $i$ ) and  $\lambda$  is a Lagrange multiplier, used to minimize possible estimation error (for detail elaboration refer to IIWIS user guide).

## 4. Results

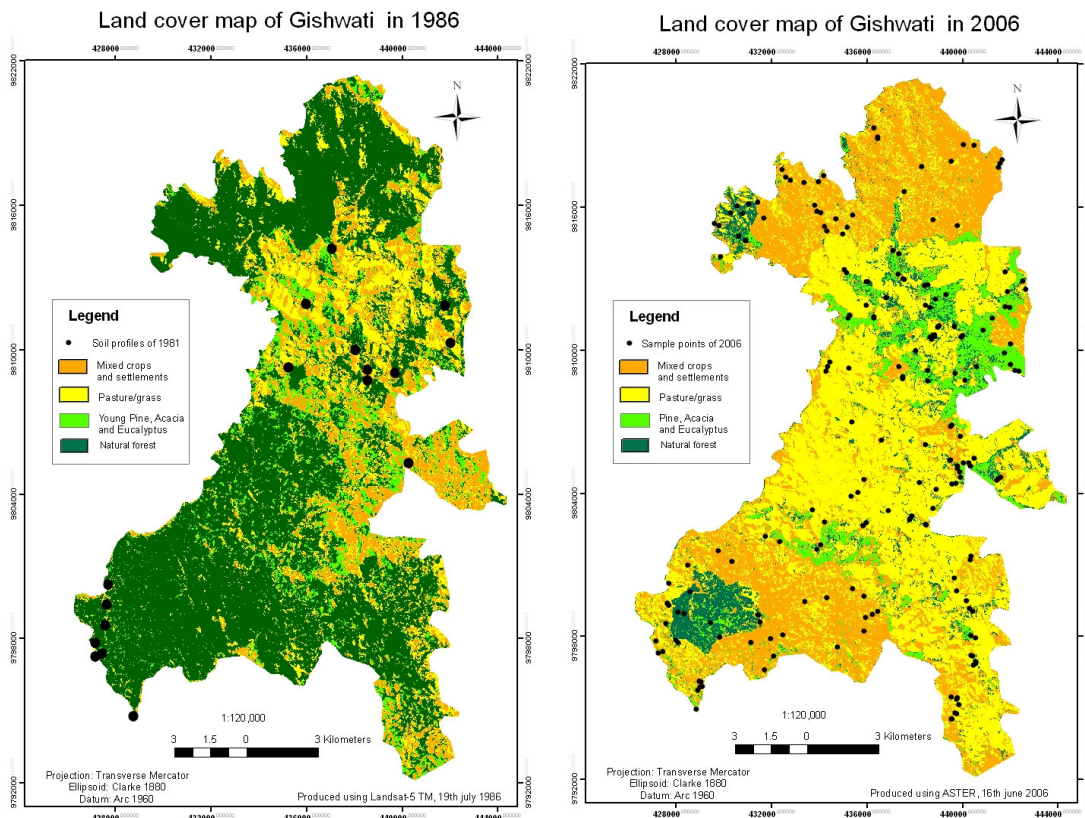
### 4.1. Characterisation of soil fertility indicators and predictors

#### 4.1.1. Land Use and Cover change (LUCC) over the 25 years and soil surveys

As shown on the Land cover maps (Figure 4-1) generated using Landsat-5 TM and ASTER, the natural forest has been exploited since 1980s. Over the years, people living in surrounding villages have often encroached upon and grazed their cattle. As reported by PCFN (Plumptre et al., 2001), about 200 km<sup>2</sup> were converted into pasture and pine plantation. The classified image showed that this conversion was done gradually. From 1986, eastern central of Gishwati was already converted to pine plantation and pasture. Based on interviews with relevant stakeholders, after 1988, the pasture land was extended to south west (Figure 4-2); about 50 km<sup>2</sup> of natural forest remained in south and 30 km<sup>2</sup> already assigned to military zone in the northern part (Fig. 4-1a). CPR (2002) reported that during soil survey, soil profiles were mostly done in the study area in 1983 during the conversion. Some profiles were done in natural forest (south), others in pasture, and young pine plantation sometimes mixture with crops (example pea nut and potatoes). Refer to the same report; the pine plantation had 3 years growth. Due to inaccessibility, only 17 soil profiles were done within the study area especially where the natural forest was already cleared (in central) and where there is accessibility to the roads (in south west) (Fig.4-1a). During that time, PENAPE project for potatoes' production and GBK project for livestock were already installed in central eastern part, which was followed by ISAR in 2000 for livestock and crops production mostly dominated by Irish potatoes.

The land cover map of 2006 (Figure 4-1b) revealed the actual status of LUC in Gishwati. Between 1995 and 2000, most of the remained natural forest and degraded areas (mostly occupied by shrubs) were converted to agriculture land and farmers were settled in south and north. To the small extent, people were settled inside the pasture land, and due to irregular encroachment the central part of pasture was then converted for crop production. Only single indigenous trees were remained in the parcels for agricultural area and those that were used to divide the paddocks of pasture remained (Fig. 4-1b).

Note that, the classified LC maps and change detection map fairly reported the land cover change and consequently the land use change in Gishwati study area because they represented accurately what was observed on the ground and what discussed with major stakeholders. The overall accuracy assessment was 83.3% for Landsat-5 TM image of July 1986 and 84.16% for ASTER image of June 2006. Change detection map was also assessed using the matrix of change and no change, and it has revealed 96.4% of accuracy.

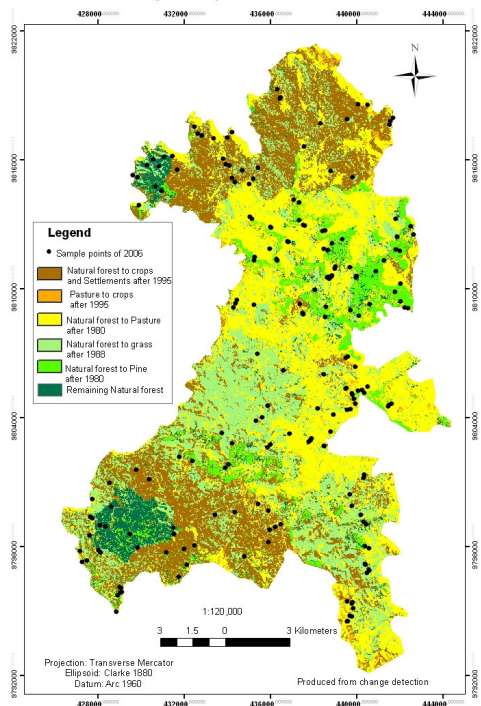


(a) Situation on 19<sup>th</sup> July, 1986

(b) Situation on 16<sup>th</sup> June, 2006

**Figure 4-1: Maps showing Land Covers of Gishwati in 1986 and 2006**

Land cover change map of Gishwati from 1980 to 2006



**Figure 4-2: Change detection map showing the change in land cover over 25 years**

Informal discussion with the farm owners in northern Gishwati (Arusha) showed that crop fields younger than 10 years since converted from the natural forest are cultivated mainly with, potatoes, maize and legumes (i.e. carrots cabbage etc.), and without fertilization due to high initial soil fertility. However, farmers in southern Gishwati (Karumbi) claimed that crop yields decline, as the fields get older unless fertilizers are applied.

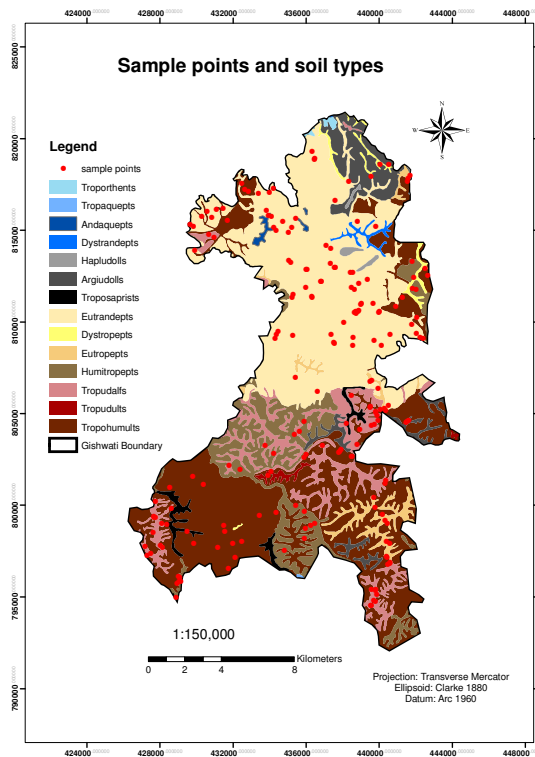
Mixed cropping as well as rotation of legumes and potatoes with cereals on the out fields is not commonly practiced in Gishwati. There was significant soil erosion in the agriculture fields (25 to 50% coverage). All the visited fields have got more or less similar management except some minor differences in the soil protection measures.

### 4.1.2. Variation of soils and soil samples locations

The map of great group soils (Figure 4-3) showed the variation of soils occurring in Gishwati study area. Eutrandepts are dominant soils in the area and lie from north to the central part of Gishwati. Those are less weathered soils developed on volcanic materials and Andaquepts occupy valleys. Dystropepts are found in east and they are developed on volcanic materials deposited on top of granites. Argiudolls are dominant in north-eastern part and are well drained soil developed on volcanic material deposited on acid material (granites). Humitropepts and Eutropepts are found in central and east and they are moderate weathered soils characterized by micas. In the south, Tropohumults and Tropudalfs are dominant soils. Thus more weathered soils are developed on mixture of acidic material (granites) and basic materials (dolerites or diorites).

As an indicator of soil fertility, out of 14 great group soils (Fig.4-3), 200 soil samples were distributed in 8 major of them. Soils that are found in small extent were not sampled. Those are Tropudults (in south), Hapludolls, Dystropepts, Trophaquepts and Troporthents (in north), and Dystrandepsts in the central of the study area.

**Table 4-1: Great group soils of Gishwati and series that are represented**



Great group	Soil Series combined
Andaquepts	MA
Argiudolls	BRI, HSA
Dystrandepsts	MAY, RUE
Dystropepts	KRO, TRE
Eutrandepts	GKB, MHR, TMA
Eutropepts	GIR, RAV
Hapludolls	GI, GO
Humitropepts	BWI, GKK, MAH, RK, RNB,
Trophaquepts	MUE
Tropodalfs	RM
Tropohumults	GSV, MUK, NBO, NBU, RAB,
Troporthents	RAM, RKO, SOV
Troposaprists	RKR
Tropudalfs	RZ
Tropudults	HNK, MB, NAR, RH
Tropudults	KAR

See appendix (2) for the description of soil series

**Figure 4-3: Great group soils map of Gishwati study area and sample locations**

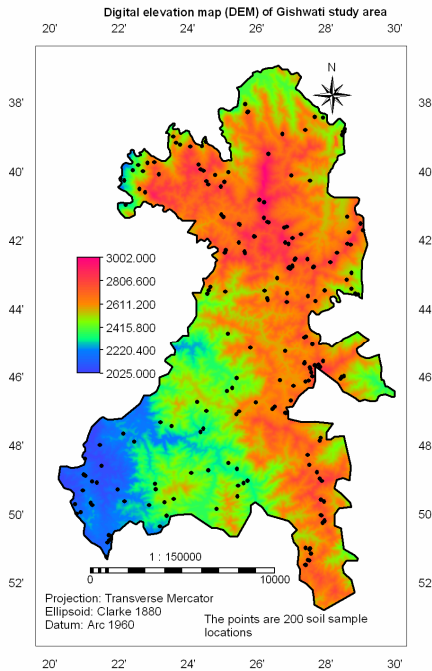
### 4.1.3. Landscape complexity of Gishwati catchment area

Gishwati catchment ranges in altitude from 2025 m to 3002 m above sea level (Figure 4-4). Muhe is known as the highest mountain in the study area which account 3003 m. The table 4-2 gives an

indication of terrain characteristics in study area. There is no distinction in use of land in term of elevation. However, at higher elevations especially on upper part of the mountain, land cover is mainly forest with *Pine* or *Acacia* in the area allocated for forest and livestock. Grassland is located on gentle slope at moderate elevation. It appears that agriculture and settlement was not previously planned to be in the area. This is shown by agriculture lands that are located in very steep part of the hills. Terraces mostly progressive and agroforestry species (*Clerodendrum: Umukuzanyana*) are used to protect the land from erosion in agriculture land especially in north and south part of Gishwati. As discussed with peoples at neighbouring of the remaining natural forest, areas under degraded natural forest are not the ones that are located in very steep part or inaccessible area, but because of the local authority stopped them to expand their activities in that area. This found to be consistent with terrain analysis. The natural forest remains in south between 2000m and 2400m whereas in the north, it is located between 2400 m and 2600 m.

**Table 4-2: Statistics' values of terrain parameters included in soil-landscape modeling**

Terrain parameters	Mean	Median	Std. Dev
DEM (m)	2541.63	2575.0	179.27
Slope (%)	36.25	35.7	20.62
SPI	924.85	901.72	531.89
STI	13.75	11.25	12.87
TWI	5.63	5.24	1.44



**Figure 4-4: Improved digital elevation map (DEM) of Gishwati catchment area**

#### 4.2. Selection of the Minimum Soil Fertility Indicators

The computation of correlations between soil properties measured from 1981 during CPR project aimed to choose minimum soil fertility indicators (MSFI) for soil fertility change mapping and modelling. As revealed (Table 4-3), the degrees of association among the 14 soil properties of the topsoil using 1300 soil samples from CPR dataset is explained by nonparametric correlation value (Spearman correlation) and the level of significant. Except for a high correlation between OC and Bulk density and porosity ( $r=0.79$ ), only low to moderate correlations were observed among physical and chemical soil fertility indicators ( $r=0.00$  to  $0.48$ ). This could be attributed to high variability of



chemical properties within the same range of physical properties in the same type of soil. High correlations were also observed among chemical soil variables such aluminum and hydrogen acidity with pH ( $r=0.52$  to  $0.81$ ). In contrast, there is low correlation between organic carbon and nutrients and exchange acidity as well. So far, the association between OC and CEC ( $r=0.37$ ) is due to the fact that in weathered soils, the source of CEC is organic matter rather than clay content. To check it, the partial correlation with controlling for Clay was computed, and revealed that out of 37 % , 21% is from OC ( $r=0.21$ ). Note that except porosity with nutrients (P, Ca, Mg), most of the associations were significant ( $P<0.05$ ).

**Table 4-3: Matrix of nonparametric correlations of 14 soil fertility indicators (SFI) in the topsoil from Rwanda (Spearman correlation: r).**

SFI	CEC	pHw	pHKCl	OC	P	Ca	Mg	K	Al	H
Clay (%)	0.17*	-0.10*	-0.08	0.34**	0.02	0.09	0.11*	0.11*	0.18*	0.11*
Silt (%)	0.27**	0.01	0.02	0.32**	-0.01	0.13*	0.12**	0.11**	0.07*	0.00
Sand (%)	-0.34**	0.04	0.00	-0.48**	-0.01	-0.20**	-0.21**	-0.18*	-0.15*	0.06*
Porosity (%)	-0.42**	-0.23*	-0.19*	<b>0.79**</b>	-0.04	0.07	0.00	0.17*	0.39**	0.12
CECcmol+.kg <sup>-1</sup>	-	-0.02	-0.01	0.37**	-0.03	0.13*	0.12*	0.10*	0.11*	0.01
pH in Water	pH	-	<b>0.89**</b>	-0.19*	0.29*	0.76**	0.76**	0.54**	<b>-0.81**</b>	-0.64**
pH in KCl		pHKCl	-	-0.13*	0.26**	0.72**	0.71**	0.52**	-0.76**	-0.64**
Organic carbon (%)			OC	-	0.09	0.09	0.68*	0.21*	0.34**	0.23**
Available Phosphorus (ppm)				P	-	0.27*	0.27*	0.33**	-0.27*	-0.22*
Exchangeable calcium (cmol+.kg <sup>-1</sup> )					Ca	-	0.92**	0.60**	-0.76**	-0.56**
Exchangeable Magnesium (cmol+.kg <sup>-1</sup> )						Mg	-	0.66**	-0.76**	0.56**
Exchangeable potassium (cmol+.kg <sup>-1</sup> )							K	-	-0.43**	0.32**
Aluminum acidity (cmol+.kg <sup>-1</sup> )								Al	-	<b>0.78**</b>
Hydrogen acidity (cmol+.kg <sup>-1</sup> )									H	-

\*\* : Correlation is significant at the 0.01 level ( $P<0.01$ )      \* : Correlation is significant at the 0.05 level ( $P<0.05$ )

Principal Component Analysis (Table 4-4) was then computed using 14 original variables and it extracted 4 major components that explain 68.9% of total variation. These showed the amount that each soil property contributes to each component. The first PC is almost a mixture of soil chemical properties except OC. The second PC has a large contribution from particle size (clay and sand), organic carbon and porosity. Yet, CEC moderately contributed to the second PC. This component largely explains the soil structure, water and nutrient holding capacity of the topsoil of Rwanda. Clay content can also explain variations in soil fertility because it may enhance the understanding of the source of acidity. In tropical rain conditions, the silicates leach to the subsurface and oxides of aluminum and other oxides (Fe, Mn) remain in the topsoil. Such soil is mostly characterized by high clay content especially Kaolinite type (1:1). In addition, sand content has a negative association to others soil variables (table 4-3). This negative relationship explains how increase of sand in topsoil decreases nutrients held in topsoil in contrast with clay content. The third and fourth PCs poorly explain variation of soil properties than the two first PCs ( $PC \text{ value} \leq 0.5$ ). PCA results meet the relationship earlier observed with Spearman correlations analysis.

**Table 4-4: Loadings of the first four components (PC) from PCA of 16 SFIs in topsoil of Rwanda**

Topsoil properties	Principal Components			
	1	2	3	4
Clay (%)	0.197	<b>0.743</b>	-0.434	0.032
Silt (%)	0.366	0.150	0.711	0.080
Sand (%)	-0.407	<b>-0.764</b>	-0.053	-0.079
pH in water	<b>0.862</b>	-0.345	0.024	0.106
pH in KCl (1N)	<b>0.803</b>	-0.331	0.115	0.133
Organic Carbon (%)	-0.036	<b>0.706</b>	0.038	0.129
Available Phosphorus (ppm)	<b>0.597</b>	-0.215	0.339	0.224
Exch. Calcium (cmol+/kg)	<b>0.761</b>	0.200	-0.335	-0.027
Exch. Magnesium (cmol+/kg)	<b>0.710</b>	0.191	-0.471	-0.103
Exch. Potassium (cmol+/kg)	<b>0.531</b>	-0.030	0.048	0.290
Aluminum acidity (cmol+/kg)	<b>-0.621</b>	0.485	0.211	0.237
Hydrogen acidity (cmol+/kg)	-0.266	-0.019	-0.235	0.850
CEC (cmol+/kg)	0.374	0.456	0.287	-0.213
Porosity (%)	0.068	<b>0.848</b>	0.223	-0.030
<b>Eigen value</b>	<b>4.050</b>	<b>3.172</b>	<b>1.396</b>	<b>1.030</b>
<b>Proportion of variance (%)</b>	<b>28.928</b>	<b>22.660</b>	<b>9.972</b>	<b>7.356</b>
<b>Cumulative proportion (%)</b>	<b>28.928</b>	<b>51.588</b>	<b>61.559</b>	<b>68.915</b>

Firstly, eleven soil attributes showing the highest component score (PCA value  $\geq 0.5$ ) for the two first principal components were subjected to soil fertility variability analysis. These soil attributes include exchangeable bases (K, Ca, and Mg) and available phosphorus for ‘nutrient dynamics’, pH, aluminum acidity (Al) for ‘solute leaching and acidification’, and particle size (clay and sand), porosity and organic carbon for ‘soil structure, air and water condition’.

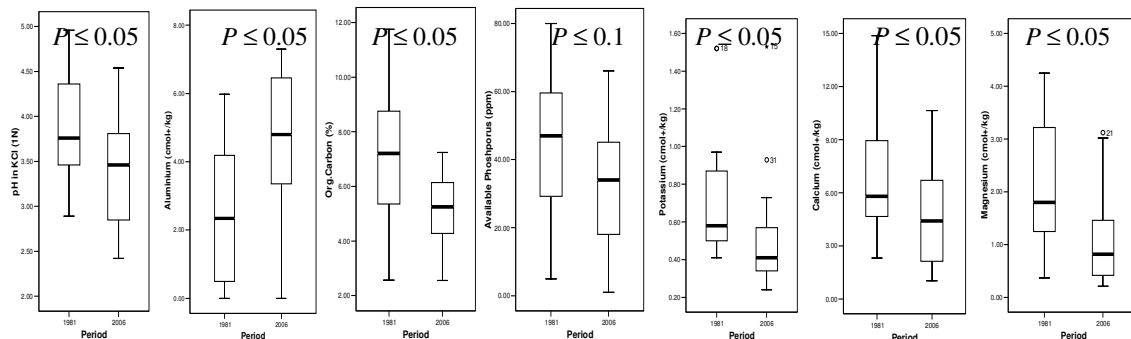
Secondly, considering economical part, organic carbon, porosity and particle size were grouped together as one indicator of soil stability because there had equal explanation to the variability (refer to second PC, PCA value range between 0.7 and 0.8). OC was chosen as representative of that group because it can satisfactory explain the soil structure in topsoil of Rwanda. It is cheaper and consumes less time in data collection compare to porosity and particle size.

Therefore, using pH, P, K, Ca, Mg, Al and OC as minimum soil fertility indicators (MSFI) would yield results similar to those using all 14 soil variables that were used in PCA, since the selected soil attributes have the highest component score within the major principal components.

### 4.3. Change in soil fertility over 25 years

The significant change was computed using Type I data of 17 samples points available in the study area. Most 17 revisited profiles were located in pasture land and pine plantation especially in area where the conversion was done since 1980s. As seen in boxplots (Figure 4-5), the variability within

each year (length of the box) and the differences between 1981 survey and 2006 measurements (median) were partially represented. In general, the soil fertility indicators changed significantly ( $P \leq 0.05$ ) as revealed by comparison of the means using one-way ANOVA (appendix 7). The average pH which 3.8 in KCl (1N) become 3.4, Al change from 2.6 to 4.5 cmol+/kg, OC (5 to 5.4%, P (42.5 to 32.2 ppm), K (0.7 to 0.5 cmol+/kg), Ca (6.7 to 4.7 cmol+/kg) and Mg (2.09 to 1.07 cmol+/kg). However there was a high variability within each MSFI (see the standard deviation values in appendix 4 and 5). This variability was confirmed by positive skewness remarked for pH and exchange bases while the negative skewness was found in organic carbon (OC), exchange acidity (Al, H) and available phosphorous (P). In addition to the differences in soil of 1981 and 2006 between different Land Uses (LU) for the revisited points, the changes in soil fertility remarked could be also explained by the variability in lithology and landscape as well. There was a similar trend in values (North-South) but with only 17 samples the change had limited explanation. Type II data was processed for detailed explanation.



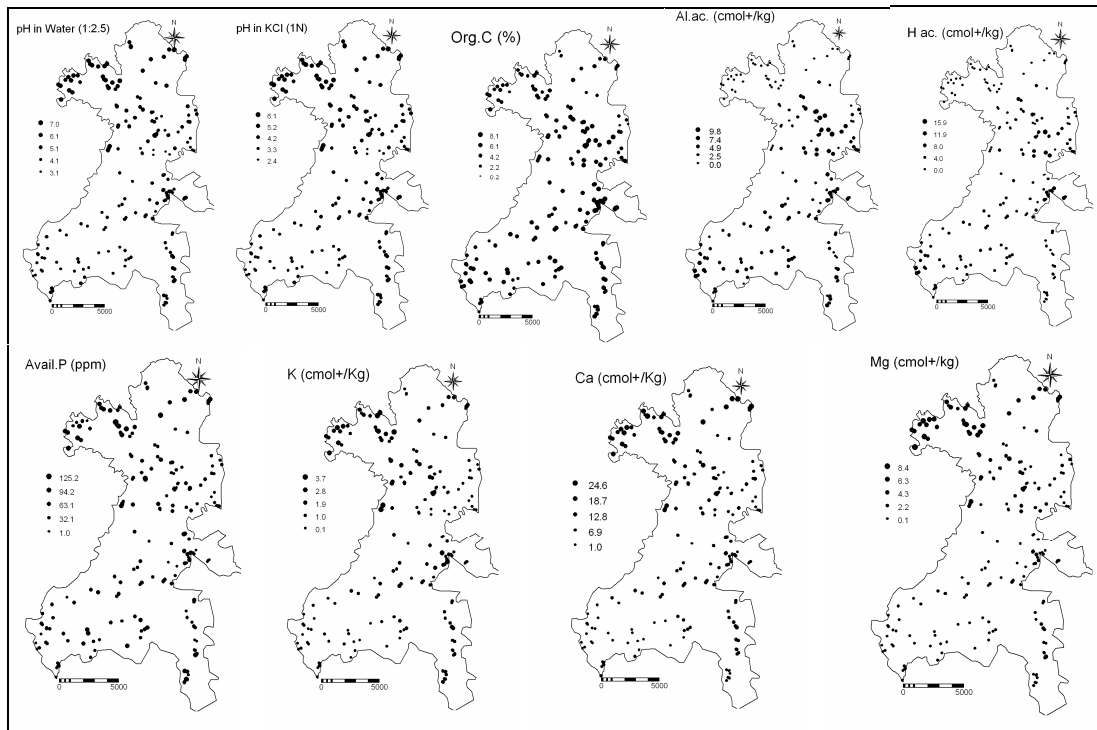
**Figure 4-5: Boxplots of each soil fertility indicator by 25 years and the probability of significant difference using one-way ANOVA.**

One-way ANOVA (Table 4-5) showed that this difference was significant ( $P < 0.05$ ). Organic carbon decreased from 7.10% to 6.5% which was considered to be a significant change. Exchange acidity almost doubled and it might affect the presence of exchangeable bases which decreased significantly especially Ca and Mg. However, there was a slightly decrease in available phosphorous. Globally, the decrease in MSFI values could be attributed to overland flow which took place after deforestation and exposition to erosion particularly in south and north part of the study area but is further explained in the next section (4.4). Refer to the standard deviation (Appendices 4 and 5); there were high variability in measured MSFIs due to high variability in soil types, Land Uses, and topography as well. This is also confirmed by further analysis (section 4.4).

#### 4.4. Spatial variation of MSFI 2006 in relation to Ancillary Variables

The actual state of soil fertility was highly variable (appendix 5) and this variability could not be explained without any reference to the main use of land, with regard to the complexity of the lithology, and topography as well. The pH in water ranged between 3.12 to 7.03, OC (0.2 to 8.1%), Al (0.0 to 9.85 cmol+/kg), P (1 to 125.23 ppm), K (0.6 to 3.7 cmol+/kg) Ca (1.04 to 24.60 cmol+/kg) and Mg (0.12 to 8.38 cmol+/kg). Except for organic carbon, the general decrease of MSFI values moved smoothly from north to south following mainly the change in soil type, secondly land

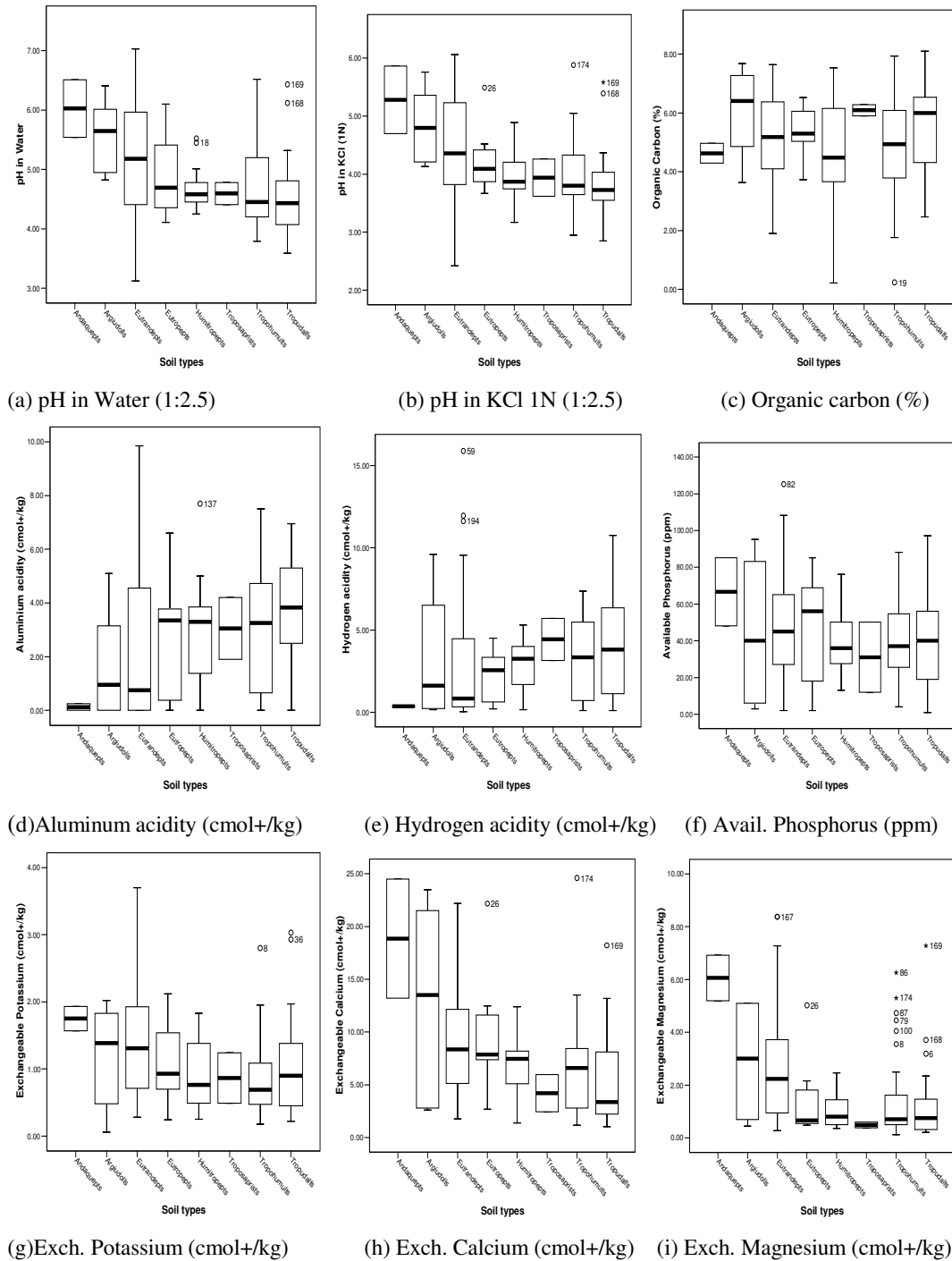
conversions, and from valley to summit as follow (Fig. 4-6); but this relationship is analysed in next sections (4.4.1- 4.4.3).



**Figure 4-6: Post plots of the minimum soil fertility indicators (MSFI) using 200 soil samples measured in the Gishwati study area**

#### 4.4.1. Variation of MSFI in relation to soil types

As shown by boxplots (Figure4-7), the decrease in nutrients (P, K, Ca, Mg) was from less weathered soils developed on volcanic materials in north (Andaquepts, Eutrandedpts) and on a mixture of volcanic and acidic material in north-east (Argiudolls) passing through the moderate weathered soils (Humitropepts and Eutropepts) and ends up to more weathered soils in the south (Tropudalfs and Tropohumults) where human activity had large influence. Tropudalfs and Tropohumults are developed on mixture of acidic material (granites) and basic materials (dolerites or diorites). However, Tropudalfs were also found in North West in volcanic material. The observations that were taken in such soils appeared in the box as unusual (e.g. nr 168 and 169). For all MSFI, there was high variability within soil type. This was shown by the position of the median lines.



**Figure 4-7: Boxplots showing MSFI variation in different type of soils**

Although the difference were highly significant for exchangeable bases ( $P < 0.01$ ), exchange acidity (Al, H), organic carbon and available phosphorus were not ( $p > 0.05$ ). They were not associated to soil types (Appendix 8). Separation of means by Duncan test distinguished five subsets of pH in relation to soil types (Table 4-5). Tropudalfs had lower pH (4.52) than others, Argiudolls and Andaquepts were the highest in pH. Although aluminium acidity was not highly different in soil types, Duncan separated its trend into three subsets. Andaquepts had lower aluminium acidity and Tropudalfs were

the highest in aluminium toxic. Exchangeable bases (Ca and Mg) were separated into 4 groups and in general, Troposapristis were the lowest loaded in cations than others (0.86, 4.21 and 0.49  $\text{cmol}+\text{kg}^{-1}$ ) respectively for K, Ca and Mg). Andaquepts were registered as the highest (1.75, 18.8 and 56.06  $\text{cmol}+\text{kg}^{-1}$ ) in bases content and followed by Argiudolls (1.19, 12.91 and 2.88  $\text{cmol}+\text{kg}^{-1}$ ).

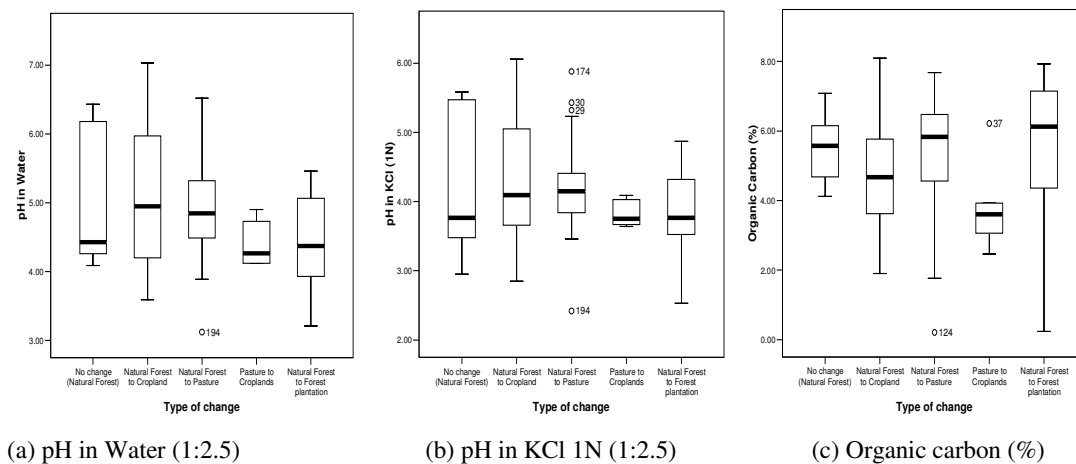
**Table 4-5: Multiple comparison of MSFI means by soil type and separations using Duncan test**

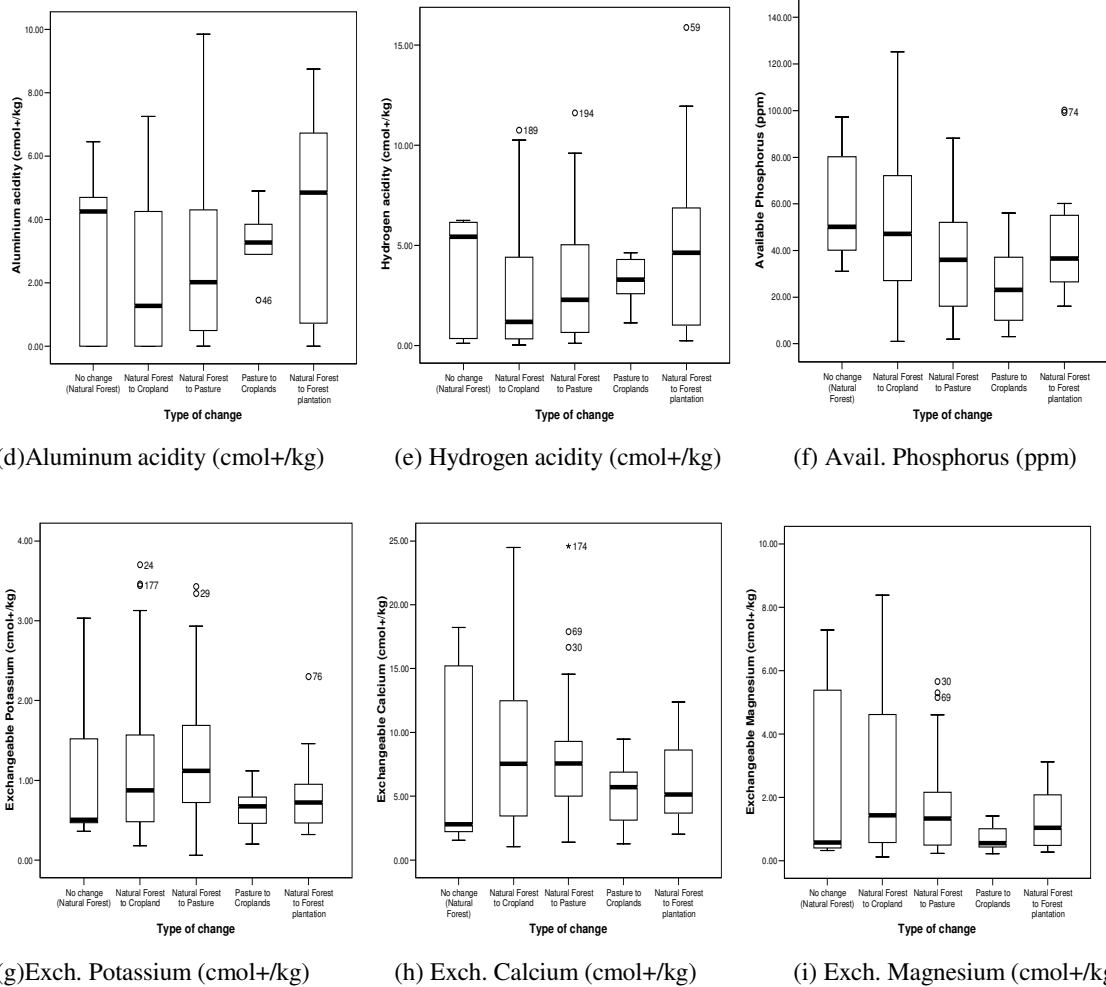
Soil Types	pH <sub>W</sub> **	pH <sub>KCl</sub> **	OC <sup>ns</sup>	Al <sup>s</sup>	H <sup>ns</sup>	P <sup>ns</sup>	K**	Ca**	Mg**
Tropudalfs	4.52 <sup>a</sup>	3.82 <sup>a</sup>	4.62 <sup>a</sup>	3.69 <sup>b</sup>	4.12 <sup>a</sup>	38.76 <sup>a</sup>	1.01 <sup>a</sup>	5.29 <sup>a</sup>	1.22 <sup>ab</sup>
Troposapristis	4.59 <sup>ab</sup>	3.94 <sup>ab</sup>	6.09 <sup>a</sup>	3.05 <sup>ab</sup>	4.42 <sup>a</sup>	31.05 <sup>a</sup>	0.86 <sup>a</sup>	4.21 <sup>a</sup>	0.49 <sup>a</sup>
Humitropepts	4.68 <sup>ab</sup>	3.98 <sup>ab</sup>	4.73 <sup>a</sup>	2.92 <sup>ab</sup>	2.92 <sup>a</sup>	40.20 <sup>a</sup>	0.91 <sup>a</sup>	6.78 <sup>ab</sup>	0.96 <sup>ab</sup>
Trophumults	4.71 <sup>ab</sup>	4.00 <sup>ab</sup>	4.85 <sup>a</sup>	2.99 <sup>ab</sup>	3.21 <sup>a</sup>	40.39 <sup>a</sup>	0.83 <sup>a</sup>	6.37 <sup>ab</sup>	1.25 <sup>ab</sup>
Eutropepts	4.91 <sup>ab</sup>	4.26 <sup>ab</sup>	5.38 <sup>a</sup>	2.60 <sup>ab</sup>	2.16 <sup>a</sup>	45.31 <sup>a</sup>	1.11 <sup>a</sup>	10.10 <sup>ab</sup>	1.55 <sup>ab</sup>
Eutrandepts	5.14 <sup>abc</sup>	4.44 <sup>abc</sup>	5.16 <sup>a</sup>	2.26 <sup>ab</sup>	2.68 <sup>a</sup>	48.29 <sup>a</sup>	1.40 <sup>a</sup>	9.41 <sup>ab</sup>	2.71 <sup>ab</sup>
Argiudolls	5.58 <sup>bc</sup>	4.84 <sup>bc</sup>	6.04 <sup>a</sup>	1.69 <sup>ab</sup>	3.28 <sup>a</sup>	44.58 <sup>a</sup>	1.19 <sup>a</sup>	12.91 <sup>b</sup>	2.88 <sup>b</sup>
Andaquepts	6.02 <sup>c</sup>	5.28 <sup>c</sup>	4.62 <sup>a</sup>	0.12 <sup>a</sup>	0.36 <sup>a</sup>	66.62 <sup>a</sup>	1.75 <sup>a</sup>	18.85 <sup>c</sup>	6.06 <sup>c</sup>

\*\* : Very significant difference ( $P \leq 0.01$ ); \* : Significant difference ( $P \leq 0.05$ ); s: Significant trend ( $P \leq 0.1$ )  
<sup>ns</sup>: No significant difference ( $P \geq 0.05$ ); <sup>a, b</sup> and <sup>c</sup> represent the subsets at  $\alpha = 0.05$ .

#### 4.4.2. Variation of MSFI in relation to Land Use change

The boxplots (Figure 4-8) showed the variability of soil fertility indicators between types of change in land use. As revealed, in contrary to exchange acidity (Al, H), the pH, organic carbon, and nutrients (P, K, Ca and Mg) decreased from no change (natural forest) to double change (pasture to cropland) passing through one change (natural forest to pasture or to cropland). For all MSFI, there was high variability within type of change. This means that land use alone is not the source of soil fertility variation in the study area. There are other factors that were also acting to soil fertility status. The variability was also explained by the skewness values for each MSFI variable (appendix 6).





**Figure 4-8: Box plots showing MSFI variation in different type of change in land use**

Multiple comparisons using one-way ANOVA (Table 4-6) showed that the type of change in Land use had a great impact on soil fertility ( $P \leq 0.05$ ). Most of the soil fertility indicators considered in this study varied across the main type of change in Gishwati study area. Duncan test showed that soils under change from pasture to cropland had lower pH and nutrients (P, K, Ca and Mg) and higher exchange acidity (Al). Soils under no change where natural forest remained and change from natural forest to croplands had lower acidity and consequently higher nutrients content. This could be explained by the time of conversion than type of land use. Refer to historical land use reported by major stakeholders and image interpretation, the conversion of natural forest being agriculture fields was done after 1995 while other areas have been converted into pasture and pine plantation since 1980. It has discovered that even though the conversion was done at the same time, pasture conserved the nutrients compare to pine plantation. Except for phosphorus, pasture had higher nutrients compared to pine plantation.

**Table 4-6: Multiple comparison of MSFI means by type of change in land use and separations using Duncan test**

Type of change	pH <sub>w</sub> **	pH <sub>KCl</sub> **	OC**	Al**	H**	P**	K*	Ca <sup>s</sup>	Mg*
Pasture to Croplands	4.40 <sup>a</sup>	3.82 <sup>a</sup>	3.81 <sup>a</sup>	3.27 <sup>a</sup>	3.20 <sup>ab</sup>	25.38 <sup>a</sup>	0.65 <sup>a</sup>	5.36 <sup>a</sup>	0.69 <sup>a</sup>
Natural Forest to Forest plantation	4.44 <sup>a</sup>	3.84 <sup>a</sup>	5.64 <sup>b</sup>	4.20 <sup>a</sup>	4.82 <sup>b</sup>	41.68 <sup>abc</sup>	0.79 <sup>a</sup>	6.28 <sup>a</sup>	1.31 <sup>ab</sup>
Natural Forest to Pasture	4.90 <sup>ab</sup>	4.19 <sup>a</sup>	5.51 <sup>b</sup>	2.54 <sup>a</sup>	2.96 <sup>ab</sup>	36.09 <sup>ab</sup>	1.24 <sup>a</sup>	7.59 <sup>a</sup>	1.59 <sup>ab</sup>
Natural Forest to Cropland	5.06 <sup>b</sup>	4.33 <sup>a</sup>	4.66 <sup>ab</sup>	2.29 <sup>a</sup>	2.45 <sup>a</sup>	50.36 <sup>bc</sup>	1.15 <sup>a</sup>	8.87 <sup>a</sup>	2.37 <sup>b</sup>
No change (Natural Forest)	5.07 <sup>b</sup>	4.29 <sup>a</sup>	5.59 <sup>b</sup>	2.90 <sup>a</sup>	3.65 <sup>ab</sup>	57.31 <sup>c</sup>	1.07 <sup>a</sup>	7.67 <sup>a</sup>	2.45 <sup>b</sup>

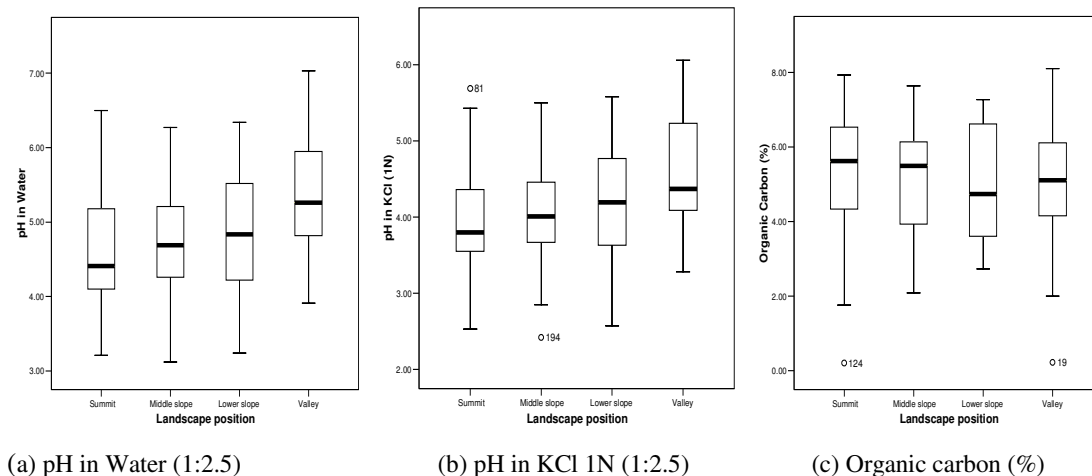
\*\* : Very significant difference ( $P \leq 0.01$ ); \* : Significant difference ( $P \leq 0.05$ ); s: Significant trend ( $P \leq 0.1$ )

<sup>a</sup>, <sup>b</sup> and <sup>c</sup> represent the subsets at groups at  $\alpha = 0.05$

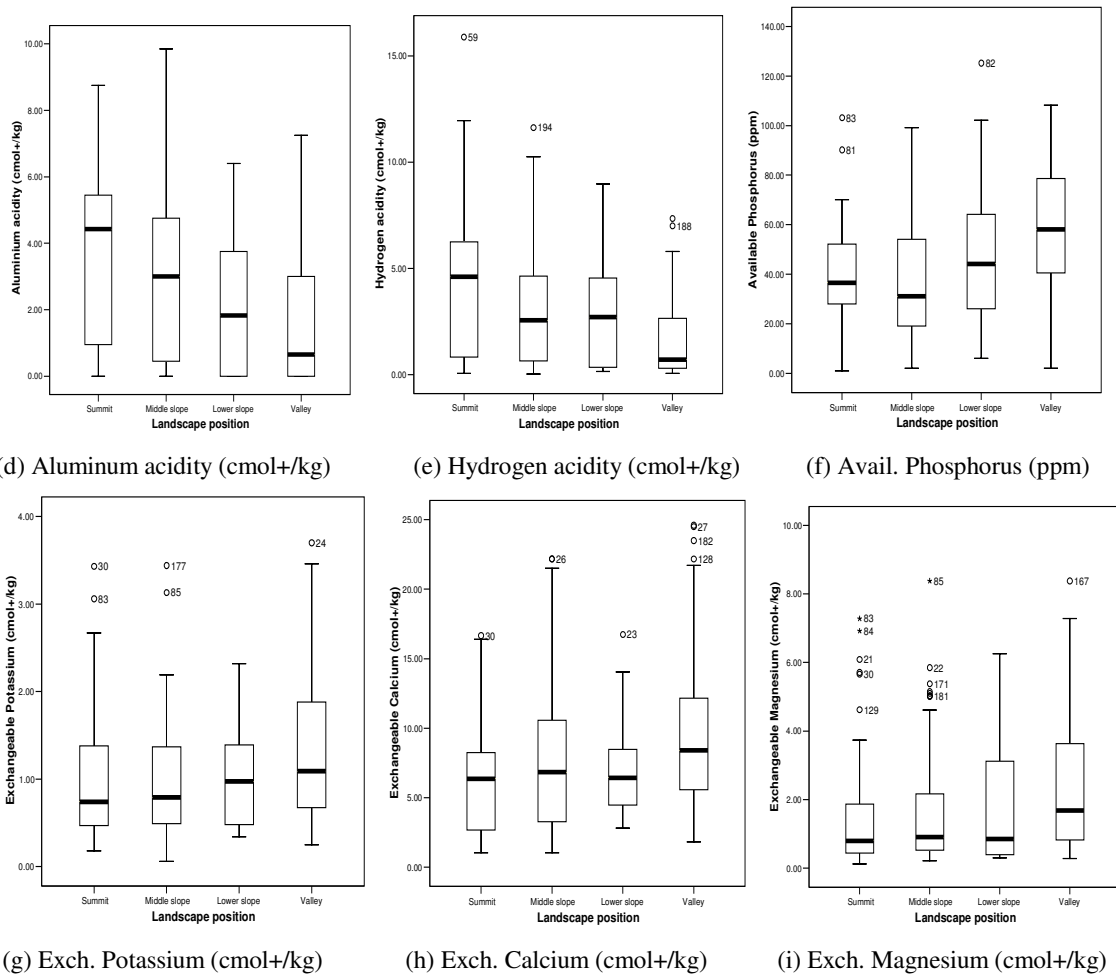
The results of analysis of MSFI by land use change revealed that even though the soil type has a great relationship with nutrients content, this relation tends to decrease when human activity is intensive. This was shown by the fact that phosphorus could not be explained by soil type (Table 4-5) but by the type of change in land use (Table 4-6). The same phenomenon was observed on exchange acidity (Al, H). The total acidity was highly associated to the type of change than soil type.

#### 4.4.3. Variation of MSFI in relation to Landscape

The pH and nutrients increased from the summit to valley for all different land uses and for all type of soils (Figure 4-9). This situation was highly remarked in agricultural area where the silicates and bases are washed away by the overland flow and oxides of Al, Fe and Mn in the surface. Most of high values were found in drainage lines and sometimes statistically appeared as unusual points.







**Figure 4-9: Soil fertility indicators for different landscape positions**

One-way ANOVA (Table 4-7) revealed that except organic carbon, other MSFI had very high significant difference between landscape positions. The lower values of pH were found on summit and higher values were found in the valleys. Soils on upper slope and summits had higher total acidity (Al, H) than soils in the valley. Nutrients (P, K, Ca and Mg) were highly concentrated in valley than upper part of the hills. Except for total acidity (Al, H) that were grouped into three groups, pH, P, and Ca were grouped into two groups of means, while K and Mg were not separated into subsets according to Duncan test.

**Table 4-7: Multiple comparison of MSFI means by landscape and separations using Duncan test**

Landscape	pH <sub>W</sub> **	pH <sub>KCl</sub> **	OC <sup>ns</sup>	Al**	H**	P**	K*	Ca**	Mg*
Summit	4.64 <sup>a</sup>	3.98 <sup>a</sup>	5.30 <sup>a</sup>	3.77 <sup>c</sup>	4.16 <sup>c</sup>	39.07 <sup>a</sup>	0.98 <sup>a</sup>	6.22 <sup>a</sup>	1.60 <sup>a</sup>
Middle slope	4.77 <sup>a</sup>	4.09 <sup>a</sup>	5.02 <sup>a</sup>	2.92 <sup>bc</sup>	3.15 <sup>ab</sup>	36.43 <sup>a</sup>	1.04 <sup>a</sup>	7.66 <sup>ab</sup>	1.70 <sup>a</sup>
Lower slope	4.89 <sup>a</sup>	4.19 <sup>a</sup>	5.07 <sup>a</sup>	2.21 <sup>ab</sup>	2.92 <sup>ab</sup>	51.31 <sup>b</sup>	1.01 <sup>a</sup>	7.63 <sup>ab</sup>	1.91 <sup>a</sup>
Valley	5.34 <sup>b</sup>	4.57 <sup>b</sup>	5.07 <sup>a</sup>	1.42 <sup>a</sup>	1.77 <sup>a</sup>	56.48 <sup>b</sup>	1.38 <sup>a</sup>	10.04 <sup>b</sup>	2.49 <sup>a</sup>

\*\* : Very significant difference ( $P \leq 0.01$ ); \* : Significant difference ( $P \leq 0.05$ ); ns: not significant difference ( $P \geq 0.05$ ); <sup>a</sup>, <sup>b</sup> and <sup>c</sup> represent the subsets at  $\alpha = 0.05$

#### 4.5. Relationships among soil fertility indicators

The spearman correlations (Table 4-8) revealed dependencies among bases cations ( $r=0.63$  to  $0.77$ ), but they were strongly affected by Aluminium acidity ( $r=-0.57$  to  $-0.73$ ). One would normally expect calcium to make up the available phosphorus but this was modified in the soils of the study area due strongly toxicity of aluminium ( $r=0.43$ ) which also explains the contribution of acidity to the complexity of P by the allophanes ( $r=-0.29$  with Al and  $-0.31$  with H). The correlation between organic carbon with total acidity was relatively strong ( $r=0.41$  with Al and  $0.42$  with H) while it had no relationship with Nutrients availability ( $r=0.08$  to  $-0.16$ ).

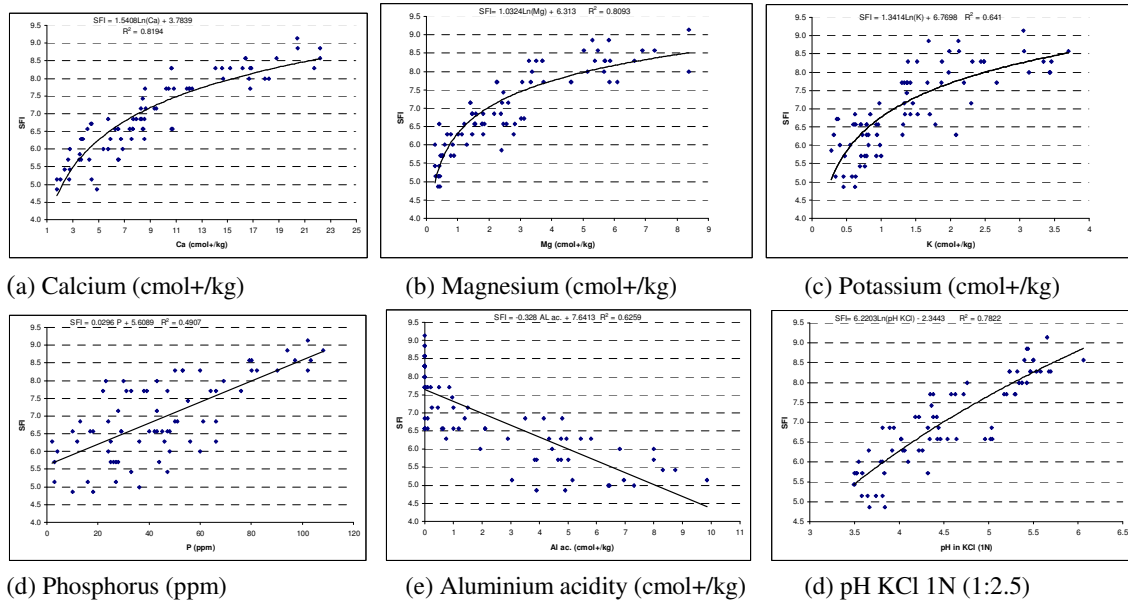
**Table 4-8: Spearman correlations among MSFI using 200 soil observations points**

MSFI	pH H2O	pH KCl	OC	Al	H	P	K	Ca	Mg
pH Water	-	0.95**	-0.23**	-0.88**	-0.78**	0.34**	0.67**	0.79**	0.73**
pH KCl 1N		-	-0.23**	-0.86**	-0.78**	0.32**	0.68**	0.80**	0.71**
Organic Carbon (%)			-	0.41**	0.42**	-0.02	0.08	-0.16*	-0.08
Aluminium acidity (cmol+/kg)				-	0.86**	-0.29*	-0.57**	-0.73**	-0.65**
Hydrogen acidity (cmol+/kg)				H	-	-0.31**	-0.58**	-0.68**	-0.60**
Available Phosphorus (ppm)					P	-	0.38**	0.43**	0.41**
Exchangeable Potassium (cmol+/kg)						K	-	0.77**	0.63**
Exchangeable Calcium (cmol+/kg)							Ca	-	0.77**
Exchangeable Magnesium (cmol+/kg)								Mg	-

\*\* Correlation is significant at the 0.01 level. \* Correlation is significant at the 0.05 level.

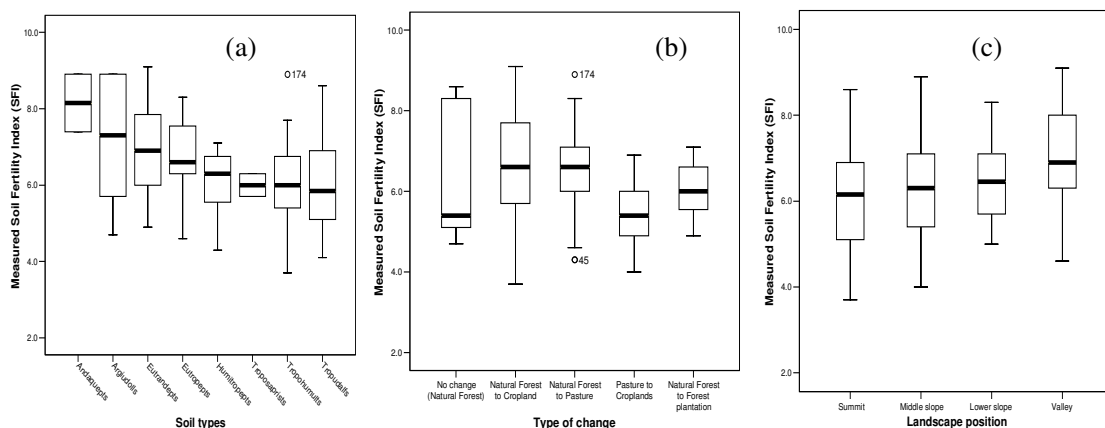
#### 4.6. Integration of Soil fertility indicators (MSFI) into index of soil quality (SFI)

After scoring MSFI in scored indicators then combined into SFI, by plotting SFI against each MSFI (Figure 4-10), the relationship between soil fertility and each soil variable was explained by the multiple determination coefficients ( $R^2$ ). The same result showed to what extent each indicator contributed to the SFI. As revealed, the exchange bases are the most contributing indicators than available phosphorous. The pH was also important than aluminium acidity (Figure 4-10e, d). The observed moderate relationship was not only due to the high variability within the indicator distribution across the study area, but also the class intervals that are assigned to the soil attribute during soil properties interpretation by Mutwewingabo et al. (1987). The fine interpretation classes, the better contribution to SFI. This constraint was specifically observed for available phosphorous (Figure 4-10d) and aluminium acidity (Figure 4-10e).



**Figure 4-10: Contribution of original MSFI values in soil fertility index (SFI)**

Comparison of soil fertility index (SFI) between soil types (Figure 4-11a) showed that SFI was significantly greater in Andaquepts than Argiudolls, Eutrandepts, and Eutropepts. The lower values were observed for Humitropepts, Troposaprists, Tropohumults and Tropudalfs. The SFI was also significant different between the type of changes. The SFI was higher where there was no change, change from natural forest to cropland, and from natural forest to pasture than for soils under change from pasture to cropland or from natural forest to forest plantation (Figure 4-11b). FI was highly significant different between landscape positions ( $P \leq 0.01$ ) and Duncan grouped SFI means in two subsets at  $\alpha = 0.05$  (table 4-9). Soil fertility increased from summit to valley (Figure 4-11c). In general SFI had the similar trend with MSFI, therefore we expect SFI to have similar spatial structure with MSFI because the continuous scored MSFI combined into SFI contains the original MSFI information of the original data.



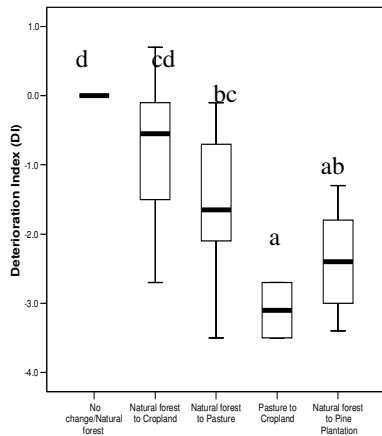
**Figure 4-11: (a) variation of SFI in different soil types, (b) type of change in land use and (c) landscape positions**

**Table 4-9: Multiple comparisons of SFI Means by soil type, land use change and landscape**

Soil type**		Land use change*		Topography**	
Great groups	SFI	Type of change	SFI	Landscape position	SFI
Tropudalfs	5.97 <sup>a</sup>	Pasture to Croplands	5.43 <sup>a</sup>	Summit	6.11 <sup>a</sup>
Troposaprists	6.00 <sup>a</sup>	Natural Forest to Forest plantation	6.06 <sup>ab</sup>	Middle slope	6.32 <sup>a</sup>
Humitropepts	6.06 <sup>a</sup>	No change (Natural Forest)	6.41 <sup>b</sup>	Lower slope	6.45 <sup>a</sup>
Tropohumults	6.07 <sup>a</sup>	Natural Forest to Pasture	6.53 <sup>b</sup>	Valley	7.03 <sup>b</sup>
Eutropepts	6.74 <sup>ab</sup>	Natural Forest to Cropland	6.62 <sup>b</sup>		
Eutrandepts	6.90 <sup>ab</sup>	***: Very significant difference ( $P \leq 0.001$ ); *: Significant difference ( $P \leq 0.05$ )			
Argiudolls	7.13 <sup>ab</sup>	a, b represent the subsets at $\alpha = 0.05$ according to Duncan test			
Andaquepts	8.15 <sup>b</sup>				

#### 4.7. Deterioration of soil fertility due to land use change

The calculated soil fertility deterioration index (DI) reflects the changes in soil fertility from Natural forest conversion to different land uses (Figure 4-12) under specific soil type. One-way ANOVA (table 4-10) revealed that soil quality in Gishwati catchment area is highly deteriorated due to land use change ( $p < 0.001$ ). Soil under double conversion from Natural forest to cultivation passing through pasture had a significantly negative deterioration ( $DI = -3.1$ ) than soils under other land use types. This area was first converted to pasture in 1980 and after 1995 the same area was converted to agriculture due to irregular encroachment in central part of Gishwati which was assigned to be grazing land.


**Table 4-10: Mean separation of DI by Duncan test**

Type of change***	Mean of DI	DI (%)
Pasture to Cropland	-3.1 <sup>a</sup>	-31
Natural forest to Pine Plantation	-2.4 <sup>ab</sup>	-24
Natural forest to Pasture	-1.6 <sup>bc</sup>	-16
Natural forest to Cropland	-0.7 <sup>cd</sup>	-7
No change/Natural forest	0.0 <sup>d</sup>	0

a, b, c, d indicate subsets at  $\alpha = 0.05$  \*\*\*: Difference between type of changes was highly significant ( $P \leq 0.001$ ).

**Figure 4-12: Deterioration index for different types of land use change**

Although the conversion of natural forest to agriculture appeared as less deteriorated compared to pasture and reforested area, this can be explained by the type of soil as well as the time of land conversion than land use type. The soils that are developed on volcanic material were slowly deteriorated compare to those that are developed on granites. Before 1995, this area was natural forest protected for military purposes (Plumptre et al., 2001). After deforestation, the area was heavily affected by erosion which accelerated the deterioration ( $DI = -0.7$ ). This is also confirmed by a positive deterioration observed in the stream. The deterioration of soil fertility for soils under grass

and reforestation with pine plantation were not the same even though the conversion was done at the same time (1980). However, deterioration is high in pine plantation (DI=-2.4) than in pasture land (DI=-1.6).

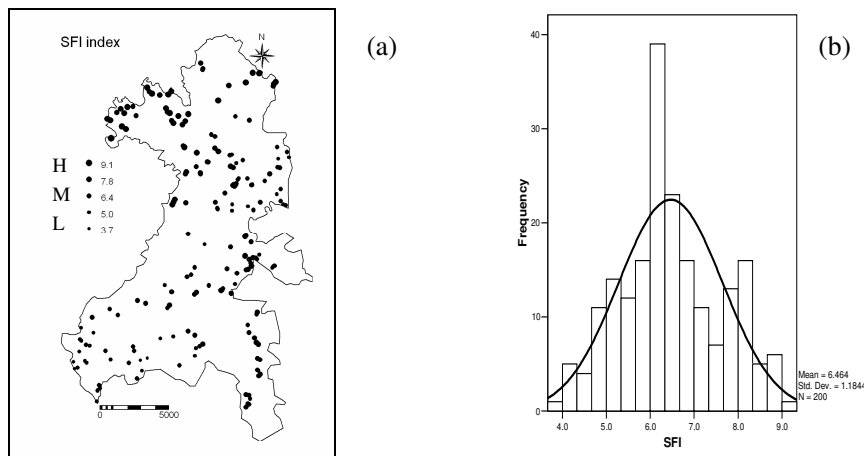
## 4.8. Geostatistical analysis of soil fertility change

### 4.8.1. Modelling the spatial structure of Soil fertility across Gishwati study area

The spatial structure soil fertility can be modeled based on the target variable, or also including covariates that are expected to influence soil properties. In this case, landscape and soil map were found as explanatory variables of soil fertility in the study area (refer to figure 4-11), so they can be used as covariates in the model soil spatial dependence.

#### 4.8.1.1. Local structure of soil fertility index (SFI)

The post plot (figure 4-13a) showed the spatial distribution of SFI. SFI exhibited a normal distribution (refer to the histogram) which resulted from the transformation of integrated soil fertility indicators into score values. This could be one of the advantages of using SFI rather than individual soil fertility indicator because it accounted the variability of indicators within the same level of soil fertility by assigning to the number of indicator values a unique score value. The score value represents the soil fertility level at sampled location (Figure 4-13).



**Figure 4-13: (a) Post plot of SFI and (b) SFI histogram using 200 sample points of 2006**

Prior to fit the model of spatial dependence to the MSFI or SFI data, spatial correlations were analyzed and have shown that pairs were correlated differently. Spatial autocorrelation coefficient of all MSFI and SFI point (Moran's value I) varied from 0.263 to 0.722.

The experimental variograms and fitted models for MSFI and SFI are shown in the figure 4-14. As revealed by the fitted exponential variogram model to pH data (Figure 4-14a), there was less unexplained variation of pH in the study area (Nug=0.1). Other variation could be explained by the

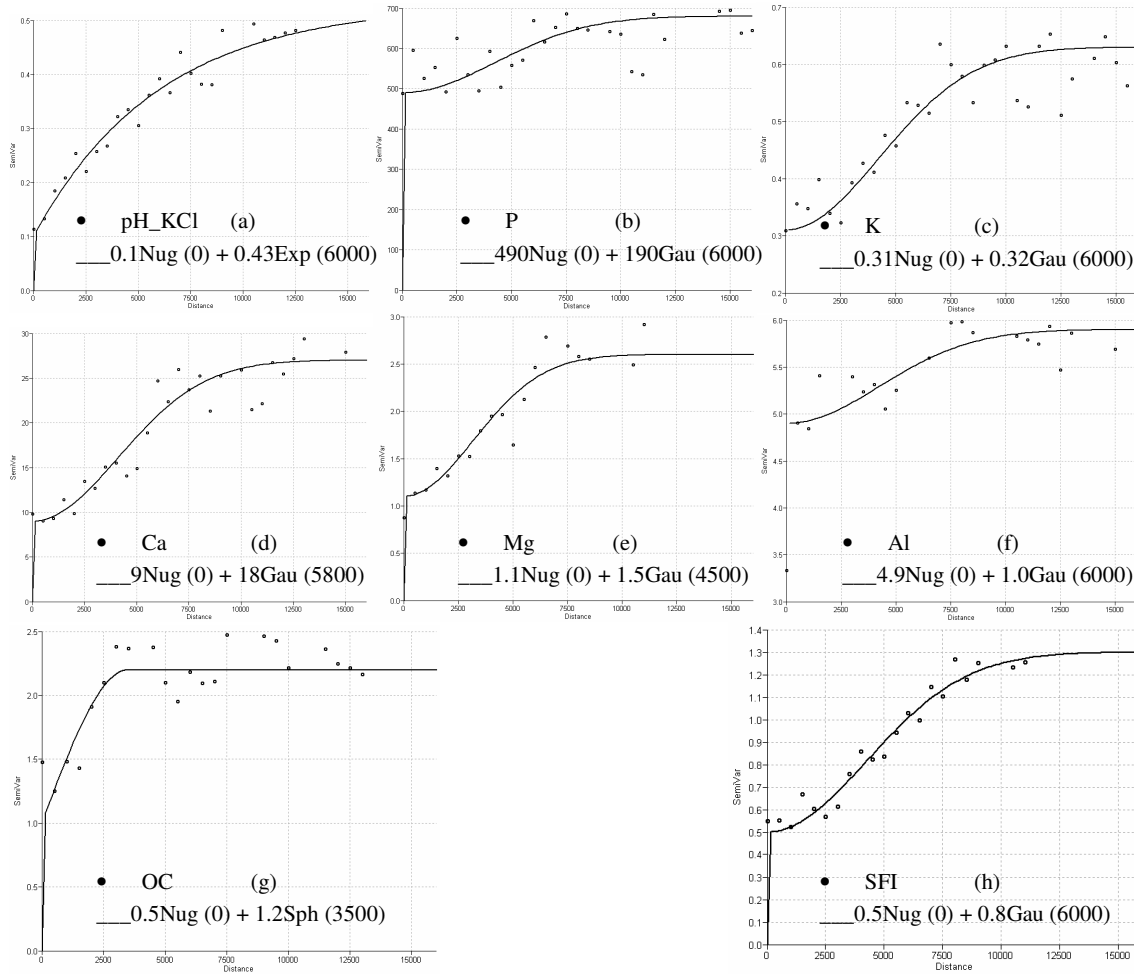
model of local structure and therefore pH was strongly explained by the local structure. This strong structure was also revealed by the high spatial correlation between pH pair points ( $I=0.568$ ). This was also observed for Base cations (K, Ca, and Mg) but they exhibited a diffusion process (Gaussian model) while pH showed an increasing spatial variation. They were spatially correlated ( $I=0.445$  for K), ( $I=0.549$  for Ca), and ( $I=0.722$  for Mg) and smaller nuggets (0.31 for K, 9 for Ca and 1.1 for Mg) indicating that the sampling scheme used resolved most of the spatial variation in base cations and pH. However, base cations yielded high residuals compare to pH (Figure 4-14c, d, and e) which meaning that the local conditions had a great impact on nutrients variability within smaller distance than on pH. The closer spatial structure observed among base cations were supported by the strong relationship observed among themselves in the previous statistical analysis ( $r=0.63$  to  $0.77$ ).

OC revealed its own spatial structure (Figure 4-14g). Spatial correlation ( $I= 0.294$ ) included in OC a short range of dependence ( $R=3.5$  km) whereas other MSFI had a large range structure (about 6 km). The short range occurred due to the historical land uses and related management, which included the division of entire area into sub-regions (refer to land cover map 2006 in the figure 4-1b) each having its own use (ether crop production, livestock or pine plantation).

Other similar cases were observed for available phosphorous (P) and aluminum acidity (Al). The higher nuggets were observed indicating very high unexplained variations in local structure (Figure 4-14e and f); and spatial correlation was lower in both elements ( $I$  value was 0.263 for P and 0.288 for Al). Such spatial behavior remarked for both P and Al confirmed the lower explanation from local ancillary variable such as lithology of the area (in this case refer to soil type). However, the latter probably reflected gradual difference in P and Al due to elevation (refer previously to landscape positions). There was very significant difference in P or Al contents from the valley floor to the mountain summits, indicating that erosion could to some extent in controlling variability process. Therefore, predicting Al or P may result the higher prediction error when compared with pH or base cations, due to the higher an explained variation in the model of spatial dependence.

The more we analyzed the local structure of each MSFI, the complex and sometimes contradictory spatial structure was revealed, indicating the weakness of each MSFI in predicting soil fertility of the study area. Some revealed the cluster at local condition (e.g. OC) or diffuse process (base cations), and others unexplained structure (Al, and P). This resulted to the transformation of original values in a probability value (SFI) that composites all behaviors into one and explain at once soil fertility of the investigated area.

The composite SFI (Figure 4-14h) yielded the spatial correlation in the middle of the Moran's value of the MSFI ( $I=0.519$ ) and showed that spatial dependence reduced after 6 km distance (refer to the range parameter of the model). A half unit of soil fertility was not explained by the local structure (refer to the nugget  $=0.5$ ). This behavior was not because of large scale drift but rather because of low variability in SFI within low distance along Gishwati catchment area. The explanatory variables such as parental material and management of the land are almost similar within smaller distance.



**Figure 4-14: Variogram models of indicators (MSFI) and resulted soil fertility model (SFI)**

As result, we chose SFI for prediction purposes and further analysis of soil fertility because it is a satisfactory representative of the MSFI both in structure and content. Secondly, it accounts the noises observed in each MSFI models. With SFI index, we expect to achieve similar and better results than the results that would be aimed using each MSFI indicator.

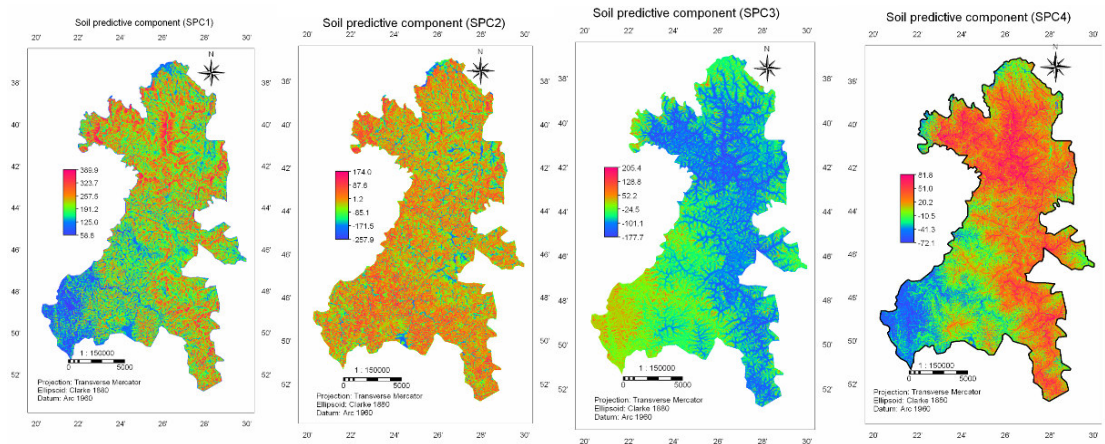
#### 4.8.1.2. Models of soil fertility structure including landscape

Detailed hillslope characterization and modelling of digital terrain and soil fertility relationships is a key to understanding the contribution of topography because they often point to the spatial distribution of significant processes such as weathering and erosion. Thus, one of our results was development of explicit landscape-based soil fertility model that can be used to improve the prediction of soil fertility along the Gishwati catena. As shown in table 4-11, DEM, slope gradient, stream power index (SPI), sediment transport index (STI) and topographic wetness index (TWI) were used to produce soil predictor component using factor analysis methods.

**Table 4-11: Factor analysis matrix of landscape predictors**

Soil predictive components (SPC)	Terrain parameters					Variance (%) per band	Cumulative variance (%)
	DEM	Slope	SPI	STI	TWI		
SPC1	0.454	0.483	0.486	0.461	0.333	74.38	74.38
SPC2	-0.185	0.372	0.367	0.007	-0.832	16.26	90.64
SPC3	-0.697	-0.019	-0.076	0.703	0.118	7.25	97.89
SPC4	0.517	0.421	-0.286	0.541	-0.425	2.08	99.87
SPC5	-0.080	-0.671	0.736	-0.024	0.041	0.03	100

The results of the factor analysis showed that there is an overlap in information and that the data can be reduced. The first four SPCs (Figure 4-15) accounted for 99.87 % of the total variation in the bands (74.38%, 16.26%, 7.25% and 2.08%). SPC1 as the main component was explained by almost equal variation in DEM, Slope, SPI, CTI and TWI. SPC2 accounted mainly for the variation in TWI, slope and SPI, while the third component accounted for DEM. As SPC1, the fourth SPC was also explained by equal mixture of DEM, STI, Slope, TWI, and SPI. Note that SPC4 revealed an opposite trend of TWI and SPI with DEM, ST, and slope compare to SPC1. The fifth accounted for Slope and SPI.



**Figure 4-15: First four soil predictive components from terrain parameters**

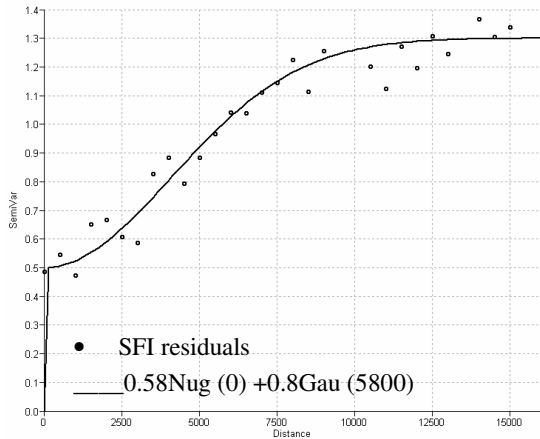
Note that factor analysis did not account only for the multicollinearity between terrain parameters but also the skewness in the original predictors.

With CLORPT approach, the more regional soil fertility behaviour was explained by multiple regression functions of soil fertility (Equation 20) derived from the two first soil predictive components (SPCs).

$$\text{SFI trend} = 5.2 + 0.006\text{SPC1} - 0.005\text{SPC2} \quad (R_a^2 = 0.12,) \quad (21)$$

Although SPCs appeared to have a weak relationship with SFI, still this association was very significant ( $P \leq 0.001$ ).





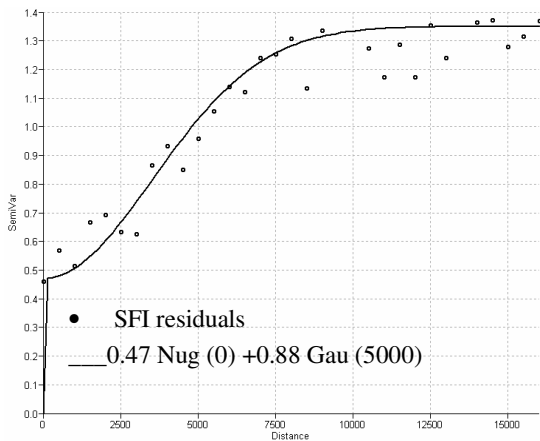
**Figure 4-16: Gaussian model of SFI residuals**

Stepwise regression analysis involved all four predictive components (SPC<sub>i</sub>) and excluded the two last components (SPC3 and SPC4) in the model. The most predictive model for SFI explained about 12 % of the SFI variation (equation 21). This model was used to interpolate the SFI trend. Then the residuals extracted from SFI trend map were modelled with geostatistics as shown in the figure 4-16.

#### 4.8.1.3. Models of soil fertility structure including both soil type and landscape

Although landscape has been considered to be strong in prediction of soil attributes, this consideration tend to decrease when there is other factors that are acting in the process. This is the case of soil fertility which not only depends on landscape but also on local lithology. Previous tests have shown that there was very high difference in SFI between soil types. Soils that are developed on volcanic material were not degraded at the same rate with soils that are developed on acidic material even though they have received the same local management.

$$\text{SFI trend} = 4.57 + 0.007\text{SPC1} - 0.006\text{SPC2} - 0.003\text{SPC3} \quad (R_a^2 = 0.25, P \leq 0.001) \quad (22)$$



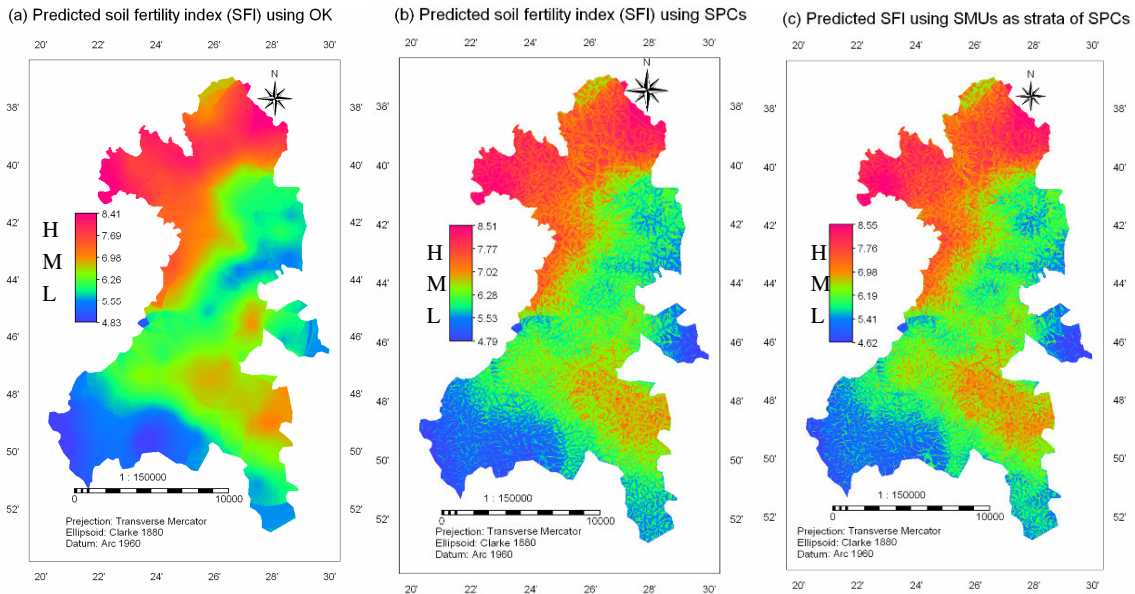
**Figure 4-17: Gaussian model of SFI residuals after stratifying SPCs by SMUs**

The contribution of Soil type in SFI was added to SFI lineal model as strata of SPCs. SPC3 which was excluded during stepwise regression analysis, was now added to the previous model. The offset was reduced from 5.2 to 4.57. As expected, this resulted to a model with high  $R_a^2$  (equation 22) and consequently less residuals (Figure 4-17). The  $R_a^2$  increased from 0.12 to 0.25. The nugget effect was also decreased from 0.58 to 0.47. In fact, Gishwati presents more or less similar landscape pattern from north to south, while its lithology is highly variable.

#### 4.8.2. Comparison of models in prediction of soil fertility

The developed models were then used to interpolate SFI point observation. The predicted SFI maps (Figure 4-18) revealed that a part from the natural environment factors that used to develop the models, the range of soil fertility classes (Low to High SFI) follows a certain event. This tempted us to attribute such pattern to different land uses in the study area. To some extent, soil type could later explain the change in the region where land use is the same. SFI predicted using SPC (Table 4-18b)

revealed details which could not be seen using smoothing Kriging (OK) or SMU map alone. After adding SMU to SPC model, SFI level was bounded in some areas (refer to blue area in NE and red area in SE of the figure 4-18c).



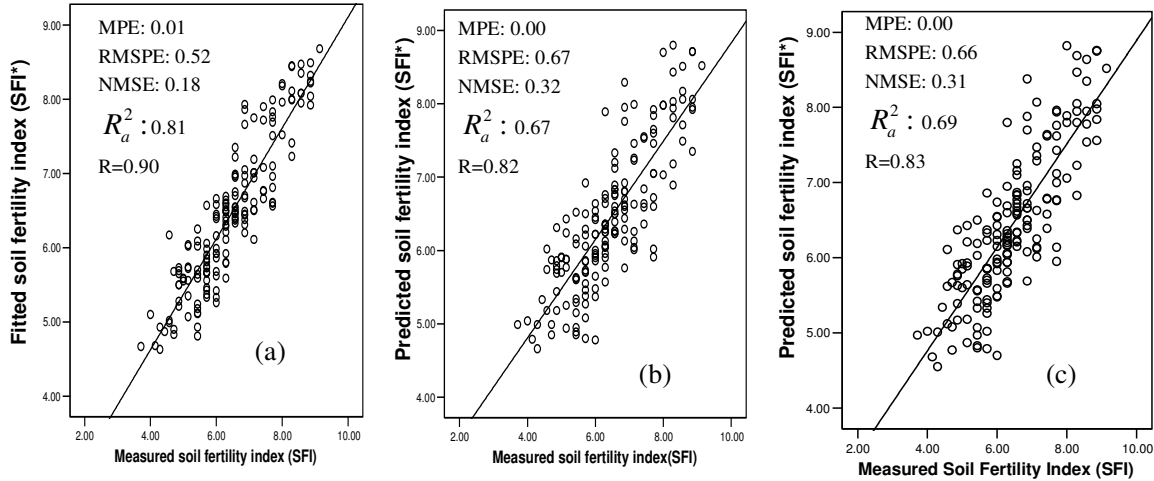
**Figure 4-18: Predicted soil fertility map (a) by OK, (b) by RK using SPCs , and (c) by RK using SMUs as strata of SPCs.**

### 4.8.3. Models quality assessment

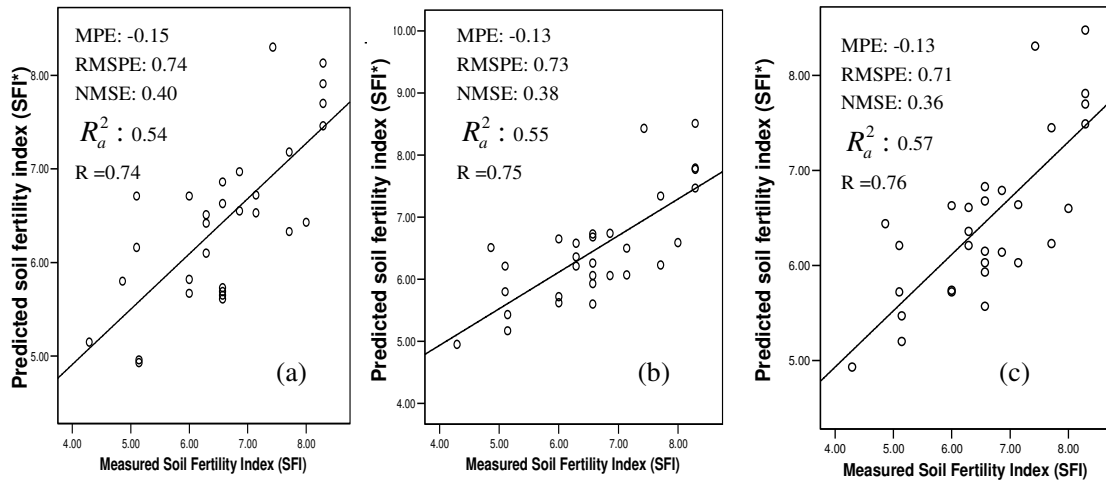
We validated the models using simple regression analysis, bias (MPE), precision (RMSPE) and relative prediction error (NMSE) in order to compare three methods used to predict SFI. The observed SFI values were regressed against those predicted ( $SFI^*$ ) from the application of the linear model combined with cross variogram model of residuals in both validation set ( $N_r=30$ ) and interpolation sets ( $N_r=170$ ). The independent data set improves the evaluation of model performance because they were not included in the model development.

The excellent correspondence between fitted and measured values for the data used to develop the models was not surprising (Figure 4-19) considering the large model value  $R_a^2$ . Whenever the same data that is perused to develop a model is also used to evaluate it, the relative prediction error (NMSE) statistic may underrepresent the prediction errors that would have been encountered by those who would use the model for prediction. Therefore, the validation with independent dataset was conducted (Figure 4-20). The relative prediction error (NMSE) with this approach was 40% by OK (Figure 4-20a), 32% by regression kriging (RK) including SPCs only (Figure 4-20b), and 31% by RK using soil type as strata of SPCs (Figure 4-20c), which was different to the error at interpolation set (18 by OK, 32% by RK using SPCs only, and 31% by RK using SPCs stratified by SMUs). This indicated that the model parameters were fairly sensitive to the individual data values used to develop the model. As expected, errors of prediction increased when the model was used to predict the values

of individual observations. The bias (MPE) occurred because all models slightly underestimated prediction soil fertility quality. The MPE was (-0.15) with OK, (-0.13) with RK using SPCs stratified by soil types (SMUs).



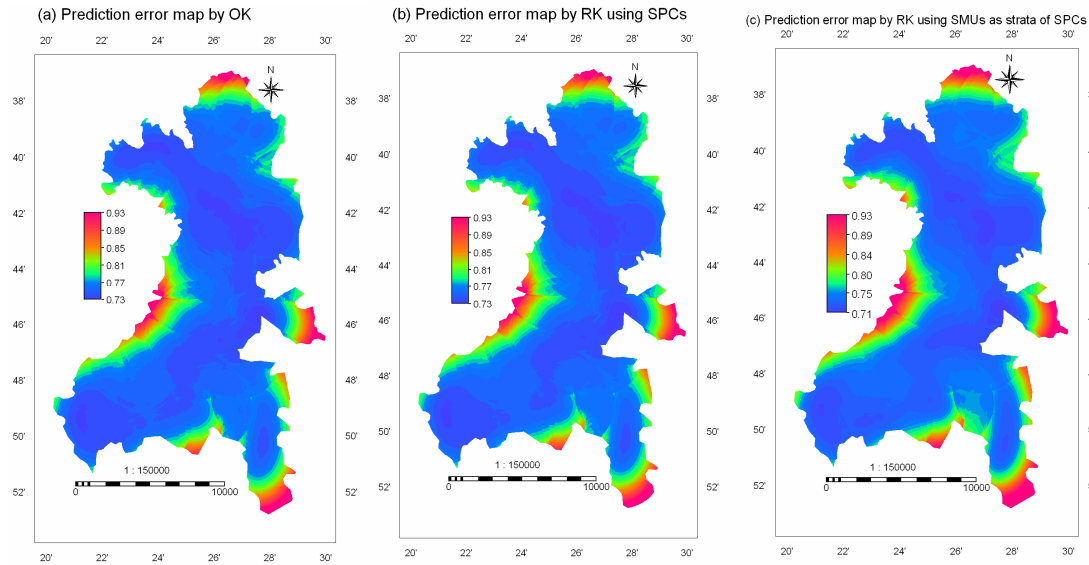
**Figure 4-19: Comparison of models performance at interpolation set (a: OK, b: RK with SPCs, and c: RK with SPCs stratified by SMUs)**



**Figure 4-20: Comparison of model performance at validation set (a: OK, b: RK with SPC, and c: RK with SMUs)**

The predicted error maps were also used to explore the distribution of bias in prediction of soil fertility of Gishwati catchment area. Refer to these maps (Figure 4-21) as the standard error or standard deviation of predicted SFI from measured SFI point, the uncertainty of prediction was estimated. As revealed, the prediction was highly accurate with RK than OK. The yielded standard error in RK is the composite error of fitted trend with stepwise regression and error that results in fitting variogram model of the residuals (Hengl et al., 2004b).

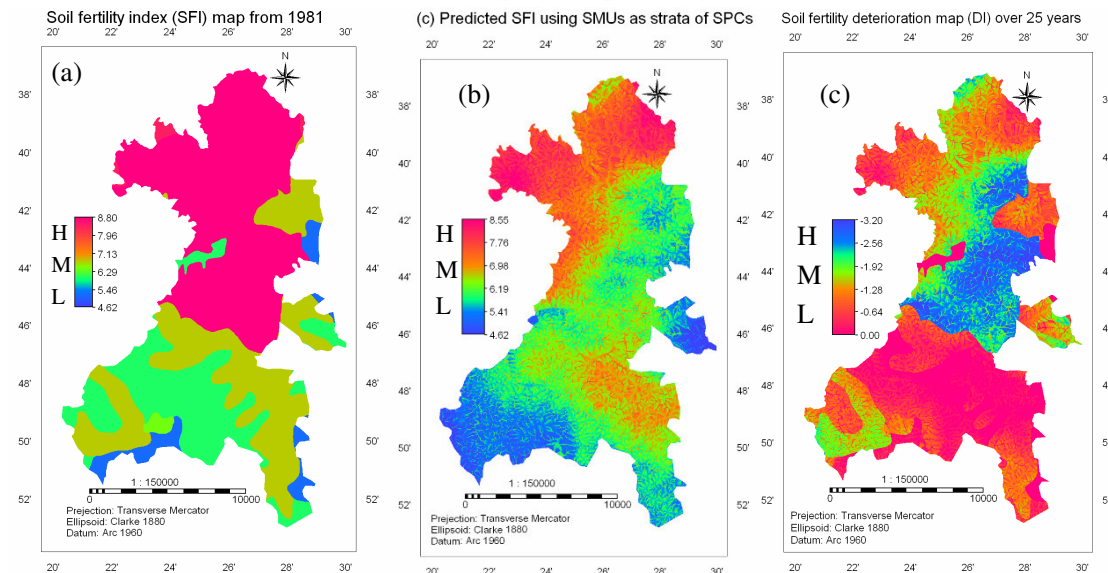
Regarding to NMSE, for all Kriging methods used, most of geostatistical parameters, the nugget and sill of spatial variability or the range of spatial correlation were fairly estimated (NMSE ≤ 40%). This was because spatial trends were associated with the order of sampling scheme which followed all possible predictors of soil fertility in the study area.



**Figure 4-21: Predicted error map (a) by OK, (b) by RK using SPC, (c) by RK using SMUs as strata of SPCs**

#### 4.8.4. Detection of the spatio-temporal soil fertility degradation

Difference map of soil fertility i.e. SFI of 1986 and SFI of 2006 revealed the general deterioration of soil fertility (DI) in Gishwati study area. As shown by Figure 4-22, DI varied from 0 to -3.2. Despite from natural event that contributed to the spread and depth of deterioration, land use change had a great influence. This was revealed by the shape of deterioration which was closer to Land use change map of the study area than other predictors. Soil type tended to have less impact on soil fertility structure in Gishwati catchment area, after human activity took place. While SFI of 1981 trend followed entirely soil type (Figure 4-22a), SFI 2006 trend deviated to structure previously drawn by soil type (Figure 4-3) by including news event such land use type (Figure 4-2). However, it was difficult to state with evidence, because compared maps were generated using different methods.



**Figure 4-22: Soil fertility index (SFI) maps for both years (1981 and 2006) and soil fertility deterioration (DI) map resulted on differences**

## 5. Discussions

### 5.1. Determination of the Minimum Soil fertility Indicators (MSFI)

Beginning with the concept of soil quality assessment as reported in several studies (Andrews et al., 2004; Andrews et al., 2002; Arshad and Martin, 2002; Boruvka et al., 2005; Rezaei et al., 2006; Stelluti et al., 1998), Soil fertility change study started with generating minimum soil fertility indicators from 1300 topsoil data of Rwanda containing 14 measured topsoil properties, using non parametric correlation analysis and principal component analysis. These techniques resulted to minimum soil fertility indicators (MSFI) formulation which contains only seven major soil chemical properties that can be used to explain major process leading to soil fertility degradation.

Considering the number of variable in the original soil dataset, and the high variability within each soil variable (Std. Dev. appendix 3), the first four PCs explained 68.9% of variation in the potential MSFI indicators. About 51.5% of the variation among all original data sets was explained by the first two PCs in the PCA. MSFI is closer to other MDS developed during several soil studies (Rezaei et al., 2006; Yemefack et al., 2006). The principal component loading matrix (Table 4-4) shows that the seven MSFI selected for soil fertility change analysis were highly weighted variables within 14 soil variables considered during the selection.

### 5.2. Trends of change in soil fertility indicators (MSFI) in relation to LUCC

The increased soil fertility degradation is related primarily to reduction of vegetation cover, decreased biogeochemical cycling and, to a lesser extent during seedbed preparation. This is reported by many researchers (Jaiyeoba, 2003; Koch and Stockfisch, 2006). All MSFI were sensitive to land use effects (Table 4-6). The long-term response of MSFI to LUCC along biosequence is found to have two phases: an initial change with clearing natural forest being pasture or replanted forest, which latter continue into the cropping without any protection measures. The similar findings were found with land clearing by burning which followed by initial cropping in southern Cameroon (Yemefack, 2005).

When natural forest is brought into other land uses, organic carbon declines (Figure 4-8c). This is simply because organic carbon is first mineralized, or lost by erosion (Wu and Tiessen, 2002). But these decreases do not have the same rate as they depend on type of change in LU. With cropping, OC is mineralized faster due to regular tillage. Unlike with cropping, with pasture (Figure 4-8c), OC is recycled due to animal wastes but since there are no records of how often and how much animal wastes are deposited, it is impossible to give an accurate estimation for the contribution from animal wastes.

Highest Al concentrations in pine and agriculture systems were observed at the 30 cm soil depth compared with pasture land (Figure 4-8a). These findings can be explained by the decrease of pH in the same systems; and therefore increases Al solubility in the surface (Lilienfein et al., 2003). In other hand, with the tropical climate, especially in western province of Rwanda, the heavy rains and its different pattern destroy soils which are uncovered. This accelerates the deterioration of silicates and their leaching involves a loss of silica in the soil and the accumulation of oxides of Al and other oxides (Fe and Mn). The increase of Al in agriculture land responds to what is often reported in the literature as leaching effect in tropical soils (Dabin, 1985; Mutwewingabo and Rutunga, 1987), and meets Yemefack (2005) findings.

The effect of pasture use on soil fertility was generally small when compared with effects of other permanent cultures like pine plantation because it preserves more nutrients (Figure 4-11b). However, the high available phosphorus in pine plantation can be attributed to the roots activity of pine than for the grass (Table 4-6). Phosphorous that are in accessible areas (in depth) is brought to the surface by roots activities. The availability of phosphorous under forest cover confirmed the conclusions from several experiments that have shown gradual improvement of P in the surface by root activity (Islam and Weil, 2000).

### **5.3. Performance of SFI model vs to MSFI models**

By weighting the minimum soil fertility indicators (MSFI) into score values, SFI index is an additive index which combine the appreciation of each MSFI in term of land productivity and stability, and it is found to be in-line with other indices reported by some researches (Andrews et al., 2003; Andrews et al., 2002; Pervanchon et al., 2005).

Of the geostatistical analysis of MSFI and SFI, SFI was the best choice for representing spatial structure of soil fertility and changes that happened due to LUCC in hilly slope condition (Figure 4-14). The relative improvement of SFI over MSFI was consistent and large. The relationship with CLORPT factors remained as it was with MSFI. However, the nugget effect was smaller while in some cases of MSFI (e.g. P, and Al) it was very high. Moreover, the experiment variogram of SFI had bounded sill, which indicates that the noise has indeed been removed.

Incorporating soil map and soil predictive components derived from DEM and derived hydrological indices in spatial prediction of SFI gives better results over OK (Figure 4-18). The range parameter of residuals was much shorter, and nugget was proportionally reduced, indicating that some variations which were not explained by the model fitted using only target variable, has been to some extent explained by soil type and topography. This largely confirms results obtained in other studies on digital soil mapping (McBratney et al., 2003) or mapping soil properties based on multivariate secondary data (Hengl et al., 2004b; Simbahan et al., 2006). The map of SFI generated with RK with SMUs and SPCs reveals the details which can not be seen on SFI map generated using OK. For instance in SE of Gishwati (between 48' and 50') Eutropepts was separated to Tropohumults while in OK they merged. Referred to SFI map values (Figure 4-18) also in previous analysis (Table 4-9), Eutropepts had moderate to high SFI value from 6.2 to 6.9 with average value of 6.7 while

Tropohumults low to moderated SFI (5.5 to 6.2) with average of 6.0. Yet, in the NE (between 42' and 44'), SFI was underestimated by OK in Eutrandepts ( $SFI \leq 5.5$ ) at moderated elevation ( $\approx 2400m$ ) while it was between 5.5 and 6.2 according to RK using SMU and SPCs. The difference remarked between SFI predicted using OK and RK concerned the differentiations of boundaries between polygons drawn by soil type and pattern of the topography. OK was found not realistic without considering other predictive factors.

#### **5.4. Degradation of soil fertility in Gishwati highlands**

By computing the deviation of SFI from Natural forest being agricultural purpose (referred in this study as DI), permanent pine plantation or pasture reveals the deterioration levels of soil fertility in Gishwati watershed. DI enables us to detect early degradation caused by different land uses in different times since the natural forest conversion (Figure 4-22c).

It is discovered that the deterioration occurs in soil quality when natural forest is degraded and converted to agriculture without the use of appropriate soil and water conservation. The same problem was also observed in Bangladesh (Islam and Weil, 2000) where soil under cultivated were high degraded over 20 years. At the same time, if natural forest is converted in forest plantation, the choice of species being planted is outmost important for soil quality conservation. If it was *Acacia* planted in large extent rather than pine, soils could not be degraded at such alarming rate (-24% refer back to table 4-10 or to the blue area in the figure 4-22c, NE from 42' to 44' where  $DI \geq -2.5$ ). This finding joints the experiment done in Bangladesh where soils under reforestation with *Acacia*, the DI was actually positive (Islam and Weil, 2000). The findings are consistent with other studies on soil fertility declines in the tropics following deforestation and conversion to agricultural lands (Lemenih et al., 2005a).

Lower degradation responses following deforestation are common observations in most Andepts (Andaquepts and Eutrandepts). For example in NE of Gishwati study area (between 40' and 42' in the map) where land conversion was done at the same time (early 1980s) with same use (pasture) but the deterioration of soil fertility was different ( $DI \leq -1.2$ ) in Eutrandepts (refer back to figure 4-3 and figure 4-22c) and ( $DI \geq -2.5$ ) in Tropohumults. We found that such indicated soils that contain allophanic minerals from volcanic eruptions can be much more resistant to degradation from cultivation than soil derived acidic material (Tropohumults) or from river deposits (Troportents and Tropaquepts). The similar results were observed with Andisols (Parfitt et al., 1997). The Andisol contained allophanes and ferrihydrite whereas the Inceptisols contained mica as the main clay mineral. The samples were taken under perennial pasture and cropped area with cropped with barley and brassica. They found that the stability of organic matter was greater in the Andisol than in the Inceptisol, and it was less likely to be affected by cropping. Allophanes and ferrihydrites appeared to have a stabilizing influence on a large part of the organic matter in the Andisol.

## 6. Conclusions and recommendations

### 6.1. Conclusions

The objective of this study was to quantify the response of soil fertility to human activity in Rwandan highlands using a case study of Gishwati catchment area. We integrated several methodological steps to provide a preliminary framework that can be used to assess and locate soil fertility degradation as one major concern of precision agriculture in Rwanda.

We made use of principal component analysis (PCA) and that was applied to the whole dataset of Rwanda in order to determine the Minimum Data Set (MDS) that can be use in soil fertility degradation assessment. Referred in this study as Minimum Soil Fertility Indicators (MSFI); seven major soil chemical properties were selected due to their higher score in two first PCs. Those are pH, OC, Al, P, K, Ca and Mg. With these findings we got the answer of the first question about the Minimum Data Set that can be used to assess the landscape-based soil fertility change in Rwandan highland.

Next, we collected 200 soil samples and we revisited 17 soil profiles done in 1981 during soil map project of Rwanda (CPR) in order to assess the early changes in soil fertility. Unfortunately, the previous profiles were few to allow us the chronosequential assessment (Type I) and state with evidence. In this case, we used biosequential or type II data to answer to the part of the second question which was raised as: if there a significant change in soil fertility over the last 25 years. One-way ANOVA revealed that each soil fertility indicator has been heavily affected by land use change over 25 years ( $P \leq 0.01$ ).

Study on how can the individual indicators of soil fertility be modeled into and integrative measure of soil fertility and fertility degradation as question 3, SFI was developed using an adapted conceptual framework after Andrews at al., 2003. This framework was made malleable using probabilistic approach and generating a score value of each class of soil properties that were already available after Mutwewingabo et al., 1987. Integration of MSFI into index of soil fertility (SFI) enabled us to reasonably interpret the complex data set with conflicting or contradictory trends due to irregular change in CLORPT factors.

This strategic method revealed that the loss of soil fertility is higher about 31% when natural forest is converted into pasture and latter continue into cropland. About 24 % of soil fertility is lost with reforestation of cleared natural forest without an appropriate species being planted. Note that pasture conserves more or less soil fertility compared to other land uses. Only 16% is lost in grassland for 25 years since natural forest has been converted. However it has discovered that soils under volcanic material are degraded slowly compared to acidic soils even though they are under same use. Only 7 % of soil fertility is lost after 11 years of regular cultivation. This comes to answer the fourth question to what extent land use change contributes to soil fertility change.



Geostatistical methods that utilized spatially correlated secondary information (RK) increased the quality of maps of SFI as compared to Ordinary kriging (OK). Both OK and RK yielded lower relative perdition error (36% to 40%). However, Regression kriging using soil predictive components (SPCs) generated from improved DEM with combination of soil type (SMUs) performed better than OK not only in terms of increasing map accuracy but also in visualization of details. In stepwise RK method relative improvement in map accuracy over OK was 4% and there was little loss of accuracy when the model was used to predict SFI at unsampled locations. SPCs was the most valuable and quantitative CLORPT factor for detailed mapping of SFI at watershed level, whereas the SMUs was used to represent parental material which varied from north to south of Gishwati watershed. With SFI map of 2006 we satisfactory answered the firth question on how successfully the spatial pattern of soil fertility can be predicted in complex lithology of Rwandan highlands. The prediction ranged from 91% to 93 % accuracy. However it should be emphasized that a SFI dataset with a fairly equal spreading of point is more appropriate for regression kriging.

## **6.2. Recommendations**

This study was considered as a preliminary quantitative assessment of soil fertility degradation and has involved secondary data unknown accuracy of the producer. We assumed that this secondary data contained uncertainties that may mask relationships with MSFI or SFI. Relying on the secondary information is risky because (i) SPCs used to generated the stepwise regression model may not be coincide exactly to the SFI point for several reasons such as GPS precision and so on, (ii) the range of soil properties classes used to develop SFI may mask the variability which could be highlighted if the range was made shorter. Those errors in the secondary information could propagate significant errors in the SFI prediction. To reduce uncertainties, we recommend updating existing soil map, including detailed geologic map and review existing topographic information by independently measured elevations, before any soil-landscape modelling and prediction.

Although CLORPT approach was used in our study, this empirical approach tends to decrease its ability in explaining the variability in the SFI model of spatial dependence. SFI Gaussian model yielded the high nugget effect still after the drifts used were removed. Adding quantitative human disturbance as drift may reduce residuals and consequently reduce unexplained variability. More work need to be done to develop a quantitative predictive component of land use as predictor of human-induced changes, and this may enhance the CLORPT approach in soil fertility change analysis.

Next to this study, more research should be devoted to these important topics, in particular validation of usefulness of SFI in decision making and implantation. The similar research should be conducted in different environment to test the transportability of SFI approach.

For further understanding of the degradation process, we suggest that long-term monitoring sites should established, regular soil samples and management aspect should be taken and stored in Master database. With such data, the model of temporal change in soil fertility can then be produced and SFI approach can be further improved for regular monitoring of the changes in soil fertility of Rwanda.

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# Appendices

**Appendix 1: Field-Data Collection Form for Soil Fertility Mapping and Modelling**

Identification of the sample site		Land Use and Land cover change				Fertilization		Erosion and conservation practices					
ID	Name of the site	X	Y	LP	PLU	LC/GC	TC	DC	FT	PF	EF	ED	ECP
1													
2													
3													
4													
5													
6													
7													
8													

**ID:** Sample identifier      **X:** Longitude (decimal degrees)      **Y:** Latitude (decimal degrees)

**LP:** Landscape position      1=Summit      2=upper slope      3=Middle slope      4=lower slope      5=Valley

**PLU:** Present Land Use: 1=Agriculture      2=Pasture      3=Forest plantation      4=Natural forest      5=Agroforestry (if dominant)

**LC/GC:** Land /Ground Cover: 1=Crops      2=Evergreen trees (>5m height)      3=Deciduous trees (>5m height)      4=Shrubs (<5m height)      5=Grasses

**TC:** Type of change      0=No change/Natural forest      1= Natural forest to cropland      2=Natural forest to Pasture      3=Pasture to Cropland      4 =Natural forest to forest plantation

**DC:** Date of Conversion (year)      **FT:** Fertilization Type: 1=organic 2=mineral (specify)      **PF:** Last period of fertilization (year)

**EF:** Erosion Features: 0= No erosion      1=Splash      2=Rill      3=Gully      4=Landslide      **ED:** Erosion density (% of coverage)

**ECP:** 0=Non erosion control      1=fosses      2=Agroforestry      3=Fallow      4=Mulching

**Appendix 2: Soil series of Gishwati**

Series' Symbols	Series' names	Soil Taxonomy, 1975	Series' Symbols	Series' names	Soil Taxonomy, 1975
BRI	Buruseli	Fine, mixed, isohyperthermic Vertic Argiudolls	NAR	Nyabikeri	Fine-loamy, mixed, isothermic Ultic Tropudalfs
BWI	Bwira	Fine, mixed, isothermic Typic Humitropepts	NBO	Nsibo	Clayey, mixed, isothermic Typic Tropohumults
GI	Gisa	Fine-loamy, mixed, isothermic, CumulicHapludolls	NBU	Nyabihu	Fine-loamy, mixed, isomesic Typic Tropohumults
GIR	Gihira	Fine-loamy, mixed, isohyperthermic Andic Eutropepts	RAB	Ramba	Clayey, mixed, isothermic Orthoxic Tropohumults
GKB	Gikombe	Ashy over cindery, isothermic Entic Eutrandedpts	RAM	Rambura	Coarse-loamy, mixed, isothermic Typic Tropohumults
GKK	Gakoko	Fine, mixed, isothermic Typic Humitropepts	RAV	Rubavu	Coarse-loamy, mixed, isothermic Andic Eutropepts
GO	Gaturo	Very-fine, montmorillonitic, isothermic, cumulic Aquic Hapludolls	RH	Ruhuha	Fine-loamy, mixed, isohyperthermic Vertic Tropudalfs
GSV	Gisovu	Clayey, mixed, isothermic Typic Tropohumults	RK	Ruko	Coarse-loamy, mixed, isohyperthermic Fluventic Humitropepts
HNK	Hanika	Fine, illitic, isothermic Ultic Tropudalfs	RKO	Rukoko	Fine-loamy, mixed, isothermic Typic Tropohumults
HSA	Hesha	Medial over loamy, isothermic Oxic Argiudolls	RKR	Rukore	Medial over fragmental, isothermic Andeptic Tropothents
KAR	Karambi	Clayey, mixed, isothermic DystropepticTropudults	RM	Rumuli	Fine, mixed, nonacid, isohyperthermic Aeric Tropaquepts
KRO	Karago	Sandy-skeletal, mixed, isomesic TypicDystropepts	RNB	Runaba	Fine, mixed, isothermic Typic Humitropepts
MA	Mukamira	Fine, mixed, isothermic Aeric Andaquepts	RUE	Rusekera	Medial, isothermic Typic Dystrandeps
MAH	Mahembe	Fine-loamy, mixed, isothermic TypicHumitropepts	RZ	Rubirizi	Dysic, Typic Troposaprists
MAY	Maya	Medial over fragmental, isothermic Typic Dystrandeps	SOV	Sovu	Clayey, mixed, isothermic Typic Tropohumults
MB	Mbarara	Fine-silty, mixed, isohyperthermic Typic Tropudalfs	TMA	Tamira	Medial, isothermic Udic Eutrandedpts
MHR	Muhabura	Medial over fragmental, isothermic Udic Eutrandedpts	TRE	Tare	Sandy-skeletal, mixed, isothermic, shallow Typic Dystropepts
MUK	Mukuku	Clayey, mixed, isothermic Typic Tropohumults	MUE	Murenge	Fine, mixed, isothermic Typic Humitropepts

Source: CPR user guide, 2002.

**Appendix 3: Descriptive Statistics of 15 soil properties of topsoil of Rwanda**

Soil variables	N	Range	Min.	Max.	Mean	Std. Dev.	Variance	Skew.	Kurtosis
Depth (cm)	1296	103	-5	98	22.80	10.887	118.522	0.741	2.167
Clay (%)	1300	86.0	2.60	88.60	35.800	15.353	235.739	0.611	0.465
Silt (%)	1300	76.4	1.90	78.30	20.401	12.367	152.952	1.295	1.927
Sand (%)	1300	91.2	0.00	91.20	43.792	17.821	317.595	-0.048	-0.380
pH in water	1296	7.78	3.42	11.20	5.321	0.883	0.780	0.601	1.101
pH in KCl	1261	10.04	0.67	10.71	4.398	0.812	0.659	0.890	3.132
Org.C (%)	1297	47.73	0.27	48.00	3.352	4.410	19.451	5.714	40.537
P (ppm)	651	485.0	0.00	485.00	20.411	48.105	2314.138	5.303	34.802
Ca (cmol+/kg)	1250	92.80	0.00	92.80	5.082	7.072	50.023	3.816	27.115
Mg (cmol+/kg)	1254	28.20	0.00	28.20	1.936	2.704	7.316	3.491	19.214
K (cmol+/kg)	1255	12.40	0.00	12.40	0.505	0.835	0.698	6.051	55.603
Al (cmol+/kg)	1095	30.45	0.00	30.45	1.811	2.289	5.240	3.218	25.528
H (cmol+/kg)	1116	61.10	0.00	61.10	0.622	3.016	9.100	12.118	189.032
CEC (cmol+/kg)	1244	114.7	1.60	116.30	18.192	13.626	185.684	2.486	9.369
Porosity (%)	170	49.07	38.07	87.13	57.959	10.454	109.297	0.314	-0.594

**Appendix 4: Summary statistics of revisited soil profiles of CPR 1981 (MSFI 1981, N=17)**

Statistics	pH	pH	Org.C	Al	H	P	K	Ca	Mg
	H2O	KCl	(%)	cmol+/kg	cmol+/kg	(ppm)	cmol+/kg	cmol+/kg	cmol+/kg
Mean	4.68	3.83	7.10	2.61	0.38	42.52	0.74	6.77	2.09
Standard Error	0.14	0.15	0.65	0.50	0.07	5.38	0.08	0.81	0.30
Median	4.72	3.76	7.21	2.34	0.40	47.00	0.58	5.80	1.80
Stand. Deviation	0.57	0.62	2.66	2.07	0.31	22.17	0.34	3.32	1.25
Sample Variance	0.33	0.39	7.10	4.28	0.09	491.3	0.12	11.04	1.57
Kurtosis	-0.73	-0.78	-0.78	-1.45	2.17	-0.92	1.69	0.65	-0.84
Skewness	-0.03	0.11	-0.04	0.16	1.26	-0.09	1.53	0.86	0.55
Range	2.00	2.07	9.19	5.98	1.20	74.96	1.11	12.56	3.88
Minimum	3.67	2.89	2.57	0.00	0.00	5.00	0.41	2.32	0.37
Maximum	5.67	4.96	11.76	5.98	1.20	79.96	1.52	14.88	4.25

**Appendix 5: Summary statistics of MSFI 2006 on the revisited MSFI 1981 (N=17)**

Statistics	pH	pH	OC	Al	H	P	K	Ca	Mg
	H2O	KCl	(%)	cmol+/kg	cmol+/kg	(ppm)	cmol+/kg	cmol+/kg	cmol+/kg
Mean	4.14	3.41	5.45	4.50	6.13	32.29	0.51	4.74	1.17
Standard Error	0.15	0.16	0.33	0.62	0.86	5.21	0.08	0.82	0.25
Median	4.15	3.46	5.27	4.80	6.25	34.06	0.41	4.42	0.82
Stand. Deviation	0.64	0.65	1.34	2.55	3.55	21.50	0.31	3.37	1.01
Sample Variance	0.41	0.43	1.81	6.48	12.60	462.3	0.10	11.36	1.02
Kurtosis	-0.51	-0.55	-0.40	-0.53	-1.02	-1.28	7.29	-0.27	-0.59
Skewness	0.01	0.23	-0.32	-0.78	-0.21	-0.02	2.55	0.86	0.90
Range	2.12	2.20	4.80	7.30	11.11	65.12	1.29	10.80	2.91
Minimum	3.12	2.42	2.56	0.00	0.51	1.00	0.24	1.04	0.21
Maximum	5.24	4.62	7.36	7.30	11.62	66.12	1.53	11.84	3.12



**Appendix 6: Summary Statistics of MSFI 2006**

Stat	pH		OC	Al	H	P	Na	K	Ca	Mg
	H2O	KCl								
Mean	4.90	4.20	5.12	2.71	3.05	43.76	0.14	1.12	7.90	1.91
Std Error	0.06	0.05	0.11	0.18	0.21	1.83	0.01	0.05	0.38	0.13
Median	4.79	4.07	5.27	2.65	2.37	41.58	0.09	0.93	7.23	1.07
Mode	4.12	3.81	5.63	0.00	0.23	56.11	0.02	0.46	8.35	1.46
Std Dev.	0.81	0.72	1.54	2.49	2.95	25.82	0.13	0.76	5.36	1.90
Sample Var.	0.66	0.52	2.37	6.18	8.73	666.6	0.02	0.58	28.7	3.62
Kurtosis	-0.59	-0.20	-0.12	-0.82	1.33	-0.15	2.22	1.31	1.01	1.27
Skewness	0.42	0.48	-0.48	0.49	1.13	0.55	1.29	1.26	1.16	1.46
Range	3.91	3.64	7.89	9.85	15.8	124.2	0.80	3.64	23.5	8.26
Minimum	3.12	2.42	0.21	0.00	0.03	1.00	0.00	0.06	1.04	0.12
Maximum	7.03	6.06	8.10	9.85	15.8	125.2	0.80	3.70	24.6	8.38
Count	200	200	200	200	200	200	200	200	200	200

**Appendix 7: One-way ANOVA showing the significant change of MSFIs over 25 years**

MSFI	pH <sub>W</sub> **	pH <sub>KCl</sub> *	OC*	Al*	H**	P <sup>s</sup>	K <sup>s</sup>	Ca*	Mg*
Fischer value	8.402	4.574	6.59	5.24	41.12	2.547	3.559	4.488	6.034
(F)									
Probability (P)	0.00	0.04	0.01	0.02	0.00	0.10	0.06	0.04	0.02

\*\* : Significant difference at  $P \leq 0.01$  \* : Significant difference at  $P \leq 0.05$  s: Significant trend  $P \leq 0.1$

**Appendix 8: One-way ANOVA of MSFI by Soil types**

Significant values	pH <sub>W</sub>	pH <sub>KCl</sub>	OC (%)	Al cmol+/kg	H cmol+/kg	P (ppm)	K cmol+/kg	Ca cmol+/kg	Mg cmol+/kg
Fischer value	4.064	5.059	1.159	1.565	1.082	0.977	3.610	5.647	7.131
Probability	0.000	0.000	0.328	0.148	0.376	0.449	0.001	0.000	0.000

**Appendix 9: One-way ANOVA of MSFI by type of change in land use**

Significant values	pH <sub>W</sub>	pH <sub>KCl</sub>	OC %	Al cmol+/kg	H cmol+/kg	P ppm	K cmol+/kg	Ca cmol+/kg	Mg cmol+/kg
Fischer value	4.048	3.008	5.595	3.473	3.703	4.782	2.428	1.773	3.389
Probability	0.004	0.019	0.000	0.009	0.006	0.001	0.049	0.103	0.010

**Appendix 10: One-way ANOVA of MSFIs by Landscape position**

Significant values	pH <sub>W</sub>	pH <sub>KCl</sub>	OC %	Al cmol+/kg	H cmol+/kg	P ppm	K cmol+/kg	Ca cmol+/kg	Mg cmol+/kg
Fischer value	6.782	5.951	0.316	8.103	5.345	6.247	2.376	3.892	1.967
Probability	0.000	0.000	0.768	0.000	0.000	0.000	0.026	0.002	0.054

**Appendix 11: Thresholds for interpretation of the pH results (after Mutwewingabo et al., 1987)**

pH	Extremely acidic	Very acidic	Moderately acidic	low acidic	Neutral
Score (max: 1)	0.2	0.4	0.6	0.8	1
PH water	3,5 - 4,2	4,2 - 5,2	5,2 - 6,2	6,2 - 6,9	6,9 - 7,6
PH KCl	3,0 - 4,0	4,0 - 5,0	5,0 - 6,0	6,0 - 6,8	6,8 - 7,2

Source: After Mutwewingabo& Rutunga, 1987

**Appendix 12: Thresholds for interpretation of Soil nutrients availability (P, K, Ca, and Mg)**

Appréciation	Extremely low	Very low	Low	Moderate	High	Very high
Score (max: 1)	0.0	0.2	0.4	0.6	0.8	1
P (ppm)	-	$\leq 3$	3 – 20	20 – 50	50 – 80	$\geq 80$
K (cmol+/kg)	-	< 0,1	0,1 – 0,2	0,2 - 0,6	0,6 – 1,2	> 1,2
Ca (cmol+/kg)	-	< 2	2 - 4	4 - 10	10 - 20	> 20
Mg (cmol+/kg)	< 0,2	0,2 - 0,5	0,5 – 1,5	1,5 - 3,0	3,0 – 8,0	> 8,0

Source: After Mutwewingabo& Rutunga, 1987

**Appendix 13: Thresholds for interpretation of the results of org. carbon and Aluminum acidity**

Organic carbon (%)	Appreciation	Score (max:1)
<0.29	Extremely low humic	0.0
0.29 – 0.58	Very low humic	0.2
0.58 – 1.16	Low humic	0.4
1.16 – 2.90	Moderate humic	0.6
2.90 – 4.64	Humic	0.8
4.64– 8.12 and above	Very humic	1
(Al ×100) /CECE	Limitation	Score (Max : 1)
$\geq 60$	High	0.2
45 - 60	moderate	0.4
30 – 45	Low	0.6
0 - 30	Very low	0.8
0.0 (not detected)	No limitation	1

Source: After Mutwewingabo& Rutunga, 1987

**Appendix 14: Spatial correlation of SFI observation points**

Distance (m)	Nr Pairs	Moran's I	Geary's C	Average Lag	Semi Variance
0	73	0.548	0.39	143.2	0.55
500	170	0.525	0.39	518	0.55
1000	262	0.582	0.37	999.3	0.52
1500	289	0.519	0.48	1501.9	0.67
2000	395	0.370	0.43	2007.5	0.6
2500	419	0.417	0.41	2513.2	0.57
3000	428	0.372	0.44	3010.6	0.61
3500	540	0.347	0.54	3517.8	0.76
4000	578	0.306	0.61	4001.8	0.86
4500	613	0.283	0.59	4496.9	0.82
5000	563	0.220	0.6	4988.5	0.84
5500	579	0.144	0.67	5511.4	0.94
6000	578	0.129	0.73	5998.4	1.03
6500	672	0.158	0.71	6496.9	1
7000	588	0.109	0.82	6998.1	1.15
7500	682	0.129	0.79	7504	1.1
8000	721	0.083	0.9	7987.8	1.27

**Appendix 15: Linear relationship between soil types and soil fertility index (SFI) ( $R^2=0.15$ )**

Soil types	Unit codes	Estimate	Std. Error	t value	Pr (> t )
Andaquepts	(SMU1)	8.145	0.781	10.42	<2e-16
Argiudolls	(SMU2)	-1.025	0.902	-1.14	0.2574
Eutrandepts	(SMU3)	-1.246	0.791	-1.58	0.1169
Eutropepts	(SMU4)	-1.409	0.886	-1.59	0.1134
Humitropepts	(SMU5)	-2.086	0.832	-2.51	0.0130
Troposapristis	(SMU6)	-2.145	1.105	-1.94	0.0537
Tropohumults	(SMU7)	-2.072	0.794	-2.61	0.0097
Tropudalfs	(SMU8)	-2.168	0.811	-2.67	0.0082